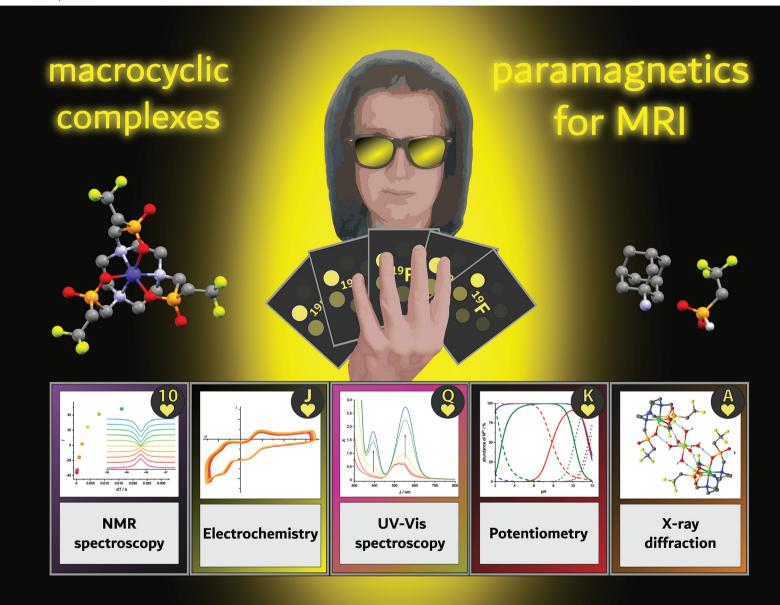
Volume 53 Number 22 14 June 2024 Pages 9227-9598

Dalton Transactions

An international journal of inorganic chemistry rsc.li/dalton



ISSN 1477-9226



PAPER

Jan Kotek *et al.*Transition metal complexes of the (2,2,2-trifluoroethyl) phosphinate NOTA analogue as potential contrast agents for ¹⁹F magnetic resonance imaging

Dalton Transactions



PAPER View Article Online
View Journal | View Issue



Cite this: *Dalton Trans.*, 2024, **53**, 9267

Transition metal complexes of the (2,2,2-trifluoroethyl)phosphinate NOTA analogue as potential contrast agents for ¹⁹F magnetic resonance imaging†

Filip Koucký, D^a Tereza Dobrovolná, D^a Jan Kotek, D*^a Ivana Císařová, Jana Havlíčková, Alan Liška, Vojtěch Kubíček D^a and Petr Hermann D^a

A new hexadentate 1,4,7-triazacyclononane-based ligand bearing three coordinating methylene-(2,2,2trifluoroethyl)phosphinate pendant arms was synthesized and its coordination behaviour towards selected divalent $(Mg^{2+}, Ca^{2+}, Mn^{2+}, Fe^{2+}, Co^{2+}, Ni^{2+}, Cu^{2+}, Zn^{2+})$ and trivalent $(Cr^{3+}, Fe^{3+}, Co^{3+})$ transition metal ions was studied. The ligand forms stable complexes with late divalent transition metal ions (from Co^{2+} to Zn^{2+}) and the complexes of these metal ions are formed above pH \sim 3. A number of complexes with divalent metal ions were structurally characterized by means of single-crystal X-ray diffraction. The complex of the larger Mn²⁺ ion adopts a twisted trigonally antiprismatic geometry with a larger coordination cavity and smaller torsion of the pendant arms, whereas the smaller ions Ni^{2+} , Cu^{2+} and Zn^{2+} form octahedral species with a smaller cavity and larger pendant arm torsion. In the case of the Co²⁺ complexes, both coordination arrangements were observed. The complexes with paramagnetic metal ions were studied from the point of view of potential utilization in ¹⁹F magnetic resonance imaging. A significant shortening of the ¹⁹F NMR longitudinal relaxation times was observed: a sub-millisecond range for complexes of Cr^{3+} , Mn^{2+} and Fe^{3+} with symmetric electronic states (t_{2g}^{3} and $HS-d^{5}$), the millisecond range for the Ni^{2+} and Cu^{2+} complexes and tens of milliseconds for the Co^{2+} complex. Such short relaxation times are consistent with a short distance between the paramagnetic metal ion and the fluorine atoms (~5.5-6.5 Å). Among the redox-active complexes $(Mn^{3+}/Mn^{2+}, Fe^{3+}/Fe^{2+}, Co^{3+}/Co^{2+}, Cu^{2+}/Cu^{+})$, the cobalt complexes show sufficient stability and a paramagnetic-diamagnetic changeover with the redox potential lying in a physiologically relevant range. Thus, the Co^{3+}/Co^{2+} complex pair can be potentially used as a smart redox-responsive contrast agent for ¹⁹F MRI.

Received 20th February 2024, Accepted 25th March 2024 DOI: 10.1039/d4dt00507d

rsc.li/dalton

Introduction

Magnetic Resonance Imaging (MRI) is one of the most powerful imaging methods in current medicine. Compared to the classical radiological methods utilizing ionizing radiation such as Computed Tomography (CT), Single-Photon Emission

Computed Tomography (SPECT) or Positron Emission Tomography (PET), MRI utilizes the nuclear magnetic resonance effect and uses non-ionizing radiation for detection which brings a strong benefit to the patients. Classical MRI utilizes the detection of the NMR effect of water ¹H nuclei and distinguishes the different tissues according to the water content and/or signal relaxation times (longitudinal, T_1 , and transversal, T_2). The image contrast can be further improved by the application of contrast agents (CAs) which influence the water proton relaxation times, and a suitable experimental setup selectively increases or decreases the water ¹H signal according to the CA distribution in the tissue. Other MRI techniques have been developed, including the detection of ¹H signals of compounds other than water (e.g. fat) or utilization of responsive ("smart") contrast agents which change their properties according to the surrounding conditions.2 In addition, other NMR active nuclei such as ¹³C, ¹⁹F or ³¹P can also be detected by MRI.3 Among them, fluorine is of special

^aDepartment of Inorganic Chemistry, Faculty of Science, Charles University, Hlavova 8, 128 42 Prague 2, Czech Republic. E-mail: modrej@natur.cuni.cz

bDepartment of Molecular Electrochemistry and Catalysis, J. Heyrovský Institute of Physical Chemistry AS CR, Dolejškova 2155/3, 182 23 Prague 8, Czech Republic † Electronic supplementary information (ESI) available: HPLC data, details on refinement of the crystal structures, discussion of the crystal structures of organic intermediates, molecular structures found in the crystal structures, selected geometric parameters, NMR spectra of the prepared compounds, UV-Vis spectra of selected complexes, distribution diagrams of the studied systems, and visualization of spectro-electrochemical experiments. CCDC 2327147–2327158. For ESI and crystallographic data in CIF or other electronic format see DOI: https://doi.org/10.1039/d4dt00507d

Paper **Dalton Transactions**

interest, as it is a monoisotopic element with a high gyromagnetic ratio which provides a very high sensitivity towards the NMR effect. The gyromagnetic ratio of ¹⁹F is very close to that of ¹H which enables the detection of the ¹⁹F NMR signal with the use of standard MRI scanners with only small hardware and software modifications. Furthermore, fluorine has almost no abundance in living organisms and, thus, it has no natural background in the imaging experiments. This fact predisposes the ¹⁹F nuclei for "hot-spot" imaging where only the externally supplied xenobiotic contrast agent molecules provide the 19F MRI signal.⁴ As both ¹H and ¹⁹F MRI experiments can be performed on the same hardware, the need to overlay 19F "hotspots" with the anatomical image can be elegantly solved by ¹H + ¹⁹F tandem MRI. ⁵ Such an imaging method could be especially useful for example for following the fate of labelled cells after transplantation.⁶

The compounds first tested as ¹⁹F NMR contrast agents were usually highly fluorinated hydrocarbons and their derivatives (for simplicity called perfluorocarbons, PFCs). The design of fluorinated contrast agents takes advantage of the generally high stability of the C-F bond towards hydrolysis which makes these compounds inert and non-biodegradable.7 Highly fluorinated organic compounds therefore usually exhibit a low acute toxicity, and some of the PFCs have been already studied for a long time as potential blood substitutes due to the high solubility of molecular oxygen⁸⁻¹¹ or as materials in vitreo-retinal surgery due to their optical properties. 12,13 Although some adverse health effects have been reported in the latter application, 14 they have been attributed mainly to the presence of trace toxic impurities like non-fully substituted hydrocarbons and other derivatives. 15,16 However, although PFCs show high stability, it was recently found that they cause some environmental risks. 17

The compounds originally used for 19F MRI are not watersoluble and have been used in the form of nano- or microemulsions which complicates some applications (e.g. cellular labelling). Further complication arises from a generally long T_1 relaxation time of the 19F nucleus in the PFC (order of seconds). Therefore, a signal is acquired for several seconds, a long delay between excitation pulses is needed and it prolongs the whole experiment. To solve this problem, complexes of fluorine-containing ligands with some paramagnetic metal ions were introduced, as the presence of a paramagnetic metal ion strongly influences the NMR relaxation times of nuclei in the vicinity. 18 The concentration of the contrast agents must be relatively high (on a millimolar scale) to achieve a reasonable signal-to-noise ratio. Therefore, the complexes must exhibit exceptional kinetic inertness as the free metal ions are frequently toxic and, in addition, the contrast agent loses convenient relaxation properties after the dissociation of the complexes. Therefore, the ligands are often based on a macrocyclic scaffold, mostly on 1,4,7-triazacyclononane (tacn), 1,4,7,10-tetraazacyclododecane (cyclen) or 1,4,8,11-tetraazacyclotetradecane (cyclam), Fig. 1, with suitably chosen coordinating pendant arms or substituents which contain fluorine atoms. The first explored groups were derivatives and analogues of cyclen-based H₄dota (Fig. 1), ¹⁹⁻³¹ the octadentate ligand family

Fig. 1 Ligands mentioned in the text.

very suitable for complexation of the lanthanide(III) ions. The Ln³⁺ complexes of such ligands are generally kinetically inert (the Gd³⁺ complexes are used as ordinary ¹H MRI CAs), ³² and a range of magnetic properties of individual Ln3+ ions brings a possibility of tuning the properties of the final CAs. 21,25,31 More recently, complexes of transition metal ions have also been found to effectively shorten the NMR relaxation times of close ¹⁹F nuclei.³³ For complexation of transition metal ions, H₄dota derivatives have also been used, ^{28,34,35} but hexadentate derivatives of tacn and cyclam are generally more suitable for this purpose.³⁶ Therefore, several tacn derivatives with three coordinating pendant arms containing fluorine were prepared and studied.³⁷⁻⁴⁰ Similarly, cyclam-based ligands with two coordinating groups have also been studied. 41-47

In our previous contributions, we showed that (2,2,2-trifluoroethyl)phosphinic acid is a suitable synthon for the introduction of the fluorine atoms into a ligand molecule. We used this coordinating pendant arm in the synthesis of the cyclenbased ligand H₄dotp^{tfe} (Fig. 1) which contains twelve fluorine atoms and we studied its Ln³⁺ complexes.³¹ Recently, the same pendant arm has been utilized in the cyclam-based ligand 1,8-H₂te2p^{tfe} (Fig. 1) containing six fluorine atoms and a study of its transition metal complexes was reported.⁴⁷ The results showed that such a concept is viable and brought promising data especially for the Co²⁺ complex.⁴⁷

To extend our work in this field, we designed a new tacn-based ligand containing three [methylene-(2,2,2-trifluoroethyl) phosphinic acid] pendant arms, H_3 notp^{tfe} (Fig. 1). It increases the number of fluorine atoms in one small molecule to nine. It would potentially increase the visibility of the contrast agents during the imaging experiment. In this manuscript, we report on the synthesis of the designed ligand and the results of its coordination study with selected first-row transition metal ions.

Experimental

Dalton Transactions

General

Commercial chemicals (Fluka, Aldrich, CheMatech, Lachema, Fluorochem) were used as obtained. Anhydrous solvents were obtained by established procedures. 48 2,2,2-Trifluoroethyl-tosylate⁴⁹ and 2,2,2-trifluoroethylbromide⁵⁰ were prepared according to published procedures with slight modifications. (2,2,2-Trifluoroethyl)phosphinic acid was prepared analogously as reported earlier⁴⁷ but 2,2,2-trifluoroethylbromide was used instead of iodide and the workup was modified. Analytical HPLC-MS experiments were carried out on a Waters ACQUITY QDa system (detection: MS - electrospray ionization under atmospheric pressure, m/z 100–1250; UV-Vis – λ 210–800 nm, diode array) equipped with an RP column (Cortecs C18 2.7 µm, 4.6 × 50 mm) using a mixture of water with 0.1% trifluoroacetic acid (TFA) and MeCN with 0.1% TFA; for details on the chromatographic method, see Fig. S1.† Mass spectra were obtained on a Waters ACQUITY QDa with a direct loading using H₂O, H₂O with 0.1% TFA or MeOH for the sample dilution. Data were processed by Empower 3 software. Flash chromatography was performed on an ECOM ECS28P0X (UV-Vis diode-array detector, λ 200–800 nm). Thin-layer chromatography (TLC) was performed on silica-coated aluminium sheets Silica gel 60 F254 (Merck). Spots were visualized using UV light (254 and 366 nm), dipping in 5% aq. solution of CuSO₄ or in aq. solution containing 1% KMnO₄ and 2% Na₂CO₃. UV-Vis spectra were recorded on spectrometer Specord 50 Plus (Analytic Jena) in a quartz-glass cell with an optical path of 1 cm. NMR spectra were recorded on a Varian VNMRS300 (frequencies 299.9 MHz for ¹H, 282.2 MHz for ¹⁹F and 121.4 MHz for ³¹P), Varian Inova 400 MHz (frequencies 400.0 MHz for ¹H, 100.6 MHz for ¹³C, 376.3 MHz for ¹⁹F and 161.9 MHz for ³¹P) or Bruker Avance III 600 MHz (frequencies 600.2 MHz for ¹H, 150.9 MHz for ¹³C and 564.7 MHz for ¹⁹F). The spectra were acquired at 25 °C unless stated otherwise. Internal references for ¹H and ¹³C NMR spectra were t-BuOH for D₂O solutions and CHD₂OD residual peak for MeOH-d₄ solutions. Aq. H₃PO₄ (3%) was used as the external reference for ³¹P NMR (0.5 ppm) and ca. 1% triflic acid for ¹⁹F NMR (-79.0 ppm). These secondary references were referenced to 85% H₃PO₄ (0.0 ppm) and to Freon-11 (0.0 ppm), respectively. Chemical shifts are given in ppm and coupling constants in Hz. Multiplicities of the signals are expressed as follows: s (singlet), d (doublet), t (triplet), q (quartet), and m (multiplet).

Chemical shifts of paramagnetic compounds were corrected for a bulk magnetic susceptibility effect as described previously (difference of the $^{19}{\rm F}$ signals of 2,2,2-trifluoroethanol between the paramagnetic sample and the insert, Table S1†); 47 the corrected values are presented throughout the text. The T_1 relaxation times of the nucleus were measured using the Inversion Recovery sequence. The $^{19}{\rm F}$ T_2 relaxation times of diamagnetic compounds were measured from the CPMG sequence on a Varian Inova 400 MHz and Bruker III 600 MHz; the measurement is not accessible on VNMRS300 as the probe used does not provide accurate 180° pulses for the $^{19}{\rm F}$ nuclei. For all paramagnetic complexes, T_2^* relaxation times were calculated from half-widths of their NMR signals.

Ligand synthesis

2,2,2-Trifluoroethyl-tosylate. 2,2,2-Trifluoroethanol (30.0 g, 300 mmol, 1.0 equiv.) was poured into a 1000 ml round-bottomed flask and was diluted with dichloromethane (DCM, 300 ml). Triethylamine (84 ml, 600 mmol, 2.0 equiv.) was added and the mixture was cooled in a bath with a water-ice mixture. To this mixture, a solution of tosylchloride (54.3 g, 285 mmol, 0.95 equiv.) in DCM (150 ml) was added dropwise. After the addition was completed, the cooling bath was removed and the reaction mixture was allowed to warm up to room temperature and was stirred for 18 h. Thereafter, ag. HCl (6 M, 150 ml) was added and the mixture was transferred into a separation funnel. The organic layer was separated, washed with brine (3 × 200 ml) and dried over anhydrous MgSO₄. Volatiles were evaporated on a vacuum rotary evaporator. The pale orange oil remaining was poured onto an evaporating dish. The white solid product crystallized in the fridge overnight. Yield 68.5 g (95%).

Elem. anal.: found (calc. for $C_9H_9F_3O_3S$, M_r 254.22) C: 42.44 (42.52), H: 3.43 (3.57), F: 21.27 (22.42), S: 12.10 (12.61).

NMR (CDCl₃): 1 H: 2.47 (s, 3H, CH₃); 4.34 (q, 2H, ${}^{3}J_{\rm HF}$ 8.0, CH₂);7.38 (d, 2H, ${}^{3}J_{\rm HH}$ 8.2, arom.); 7.81 (d, 2H, ${}^{3}J_{\rm HH}$ 8.2, arom.). 13 C{ 1 H}: 21.8 (CH₃); 64.7 (q, ${}^{2}J_{\rm CF}$ 38.1, CH₂); 122.7 (q, ${}^{1}J_{\rm CF}$ 277, CF₃); 128.2, 130.3, 132.0, 146.1 (4 × arom.). 19 F: -74.2 (t, ${}^{3}J_{\rm FH}$ 8.0, CF₃).

Recrystallization from boiling 96% aq. EtOH afforded single-crystals suitable for determination of the crystal structure by X-ray diffraction. The crystal structure is the same as that already reported, ⁵¹ but with significantly better parameters of refinement (see the ESI, Table S4 and Fig. S34†).

2,2,2-Trifluoroethylbromide. The non-trivial apparatus is shown in Fig. S2.† 2,2,2-Trifluoroethyl-tosylate (105 g, 413 mmol), diethyleneglycol (250 ml), KBr (75.0 g, 630 mmol, 1.5 equiv.) and a few ceramic boiling chips were added into a 1000 ml three-necked round-bottomed flask in a heating mantle. The flask necks were equipped with a stopcock connected to a N_2 (g) source, a Dimroth condenser and a stopper. The Dimroth condenser was connected to an adapter with a thermometer and a low-temperature side condenser which was filled with ethanol. The temperature in the low-temperature condenser was maintained around -70 °C by periodic addition of liquid N_2 . A 100 ml round-bottomed receiving flask was con-

nected to the low-temperature condenser and cooled in a Dewar vessel which was filled with ethanol and N_2 (l). Water for the Dimroth condenser was pre-heated to 35 °C (the boiling point of the product is approx. 26 °C). The reaction mixture was heated to reflux diethyleneglycol (245 °C) and 2,2,2-trifluoroethyl-tosylate and KBr fully dissolved at the high temperature. The product was distilled as a colourless high-

NMR (20% v:v solution in CH₂Cl₂, 20 °C): 1 H: 3.72 (q, 2H, ${}^{3}J_{HF}$ 8.9, CH₂). 13 C{ 1 H}: 26.4 (q, ${}^{2}J_{CF}$ 37.8, CH₂); 123.7 (q, ${}^{1}J_{CF}$ 274, CF₃). 19 F: -69.6 (t, ${}^{3}J_{FH}$ 8.9, CF₃).

density liquid. The yield was 65.9 g (98%).

(2,2,2-Trifluoroethyl)phosphinic acid and ammonium salt. Under a stream of argon, crystalline hypophosphorous acid (5.00 g, 75.8 mmol) in a three-necked 250 ml flask was dissolved in anhydrous DCM (50 ml). The flask was immersed in a water–ice cooling bath and anhydrous N,N-diisopropyl-ethylamine (DIPEA, 29.4 g, 227 mmol, 3.0 equiv.) was added. Afterwards, trimethylsilyl chloride (TMS-Cl, 24.7 g, 227 mmol, 3.0 equiv.) was added dropwise; the rate of addition was kept slow to prevent a white haze from flowing away from the apparatus. After addition of the whole amount of TMS-Cl, the reaction mixture was stirred at room temperature for 3 h. Formation of bis(trimethylsilyloxy)phosphine was confirmed by 31 P NMR (141 ppm, d, 1 J_{PH} 177).

Then, 2,2,2-trifluoroethylbromide (13.6 g, 83.5 mmol, 1.1 equiv.) dissolved in anhydrous DCM (20 ml) was added dropwise to the reaction mixture; the rate of the addition was slow enough to prevent haze formation. The mixture was stirred at room temperature overnight. Then, the reaction mixture (0.1 ml) was hydrolysed in 96% EtOH (1 ml) and ³¹P NMR of the hydrolysed sample revealed ca. 72% conversion; the main impurity was H₂PO₃. The whole reaction mixture was hydrolysed by dropwise addition of 96% EtOH (60 ml). The hydrolysed mixture was evaporated on a rotary evaporator at the bath temperature of 50 °C. The residue was dissolved in water (30 ml) and the solution was poured onto a column of a strong cation exchange resin (Dowex 50, H⁺-cycle, 400 ml of aq. suspension) and a crude product was eluted off with water. During this procedure, only a part of DIPEA was kept on the column and some DIPEA was still present in the product (1H NMR). Therefore, chromatography on the same cation exchange column (re-converted to the H⁺-cycle) was repeated three more times (until the signals of DIPEA disappeared). The aqueous solution of the purified product was concentrated on a rotary evaporator at a maximum temperature of 40 °C in the

The crude product free from DIPEA was purified on a silica column (100 g of dry SiO₂) using conc. aq. NH₃:EtOH at 1:25 as an eluent. Fractions containing the pure product were combined and evaporated in a vacuum affording ammonium (2,2,2-trifluoroethyl)phosphinate as a white solid. Yield 7.1 g (57%). Spectroscopic characteristics were identical to those reported previously.⁴⁷ The single crystals of the product were prepared by the slow evaporation of the aqueous solution to near dryness. Besides the simple ammonium salt, the single crystals of 1-adamantylammonium salt were prepared analo-

gously by evaporation of the aqueous solution of (2,2,2-trifluor-oethyl)phosphinic acid obtained from the ammonium salt by elution through a strong cation exchanger (Dowex 50, H⁺-form, elution with water) neutralized with 1-adamantylamine. For data of these crystal structures, see the ESI (Table S4 and Fig. S35, S36†).

1,4,7-Tris{[2,2,2-trifluoroethyl(hydroxy)phosphoryl]methyl}-1,4,7-triazacyclononane, H_3 notp^{tfe} (H_3 L). The ammonium salt of (2,2,2-trifluoroethyl)phosphinic acid (2.25 g, 13.6 mmol, 3.4 equiv.) was dissolved in distilled water (15 ml). The solution was loaded onto a column of a strong cation exchanger (Dowex 50, H^+ -form, 100 ml) and the free acid was eluted with water. The acid fraction was evaporated *in vacuo*. The absence of ammonium cations was confirmed by Nessler's reagent.

The oily residue was diluted in aq. TFA (50% v/v, 50 ml) and tacn (0.52 g, 4.0 mmol, 1.0 equiv.) was added. After its dissolution, paraformaldehyde (0.48 g, 16 mmol, 4.0 equiv.) was added in one portion. The flask was tightly closed and the suspension was stirred at 40 °C for 1 d. Then, an additional amount of paraformaldehyde was added (0.12 g, 4 mmol, 1 equiv.) and the reaction mixture was stirred at 40 °C for the next 1 d. After cooling, volatiles were removed in vacuum affording a brownish oil. It was diluted with water (10 ml) and loaded onto a column of a strong cation exchanger (Dowex 50, H⁺-form, 400 ml). Elution with water afforded the required product in a mixture with simple acids [starting with (2,2,2-trifluoroethyl)phosphinic acid, hydroxymethyl-(2,2,2-trifluoroethyl)phosphinic acid, (2,2,2-trifluoroethyl)phosphonic acid and traces of TFA]; the doubly substituted macrocycle derivatives remained on the column. The acid fraction was evaporated on a vacuum rotary evaporator. The crude oily product was diluted with water (10 ml) and purified in two portions by flash chromatography using a C18-AQ stationary phase (PuriFlash, 120 g). A gradient of MeCN in H₂O was used (both solvents with 0.1% of TFA) with a flow of 50 ml min⁻¹: 0.0 min 0% MeCN, 3.0 min 0% MeCN, 12.0 min 20% MeCN, 15.0 min 20% MeCN, 15.5 min 100% MeCN, and 18.5 min 100% MeCN. The product retention time was 9.8 min. The pure productcontaining fractions were evaporated to dryness and the product was isolated as a non-stoichiometric trifluoroacetate (ca. 0.3 molar equiv. according to 19F NMR). The yield was 1.26 g (ca. 49%) of colourless glassy oil.

NMR (H₃notp^{tfe}·0.3TFA, D₂O, pD 0.5): ¹H: 2.88 (dq, 6H, ² J_{HP} 14.6, ³ J_{HF} 11.8, PC H_2 CF₃); 3.44 (d, 6H, ² J_{HP} 6.4, NC H_2 P); 3.61 (s, 12H, NC H_2 CH₂N). ¹³C{¹H}: 36.5 (dq, ¹ J_{CP} 86.4, ² J_{CF} 28.4, PC H_2 CF₃); 52.4 (s, NC H_2 CH₂N); 56.3 (d, ¹ J_{CP} 99.0, NC H_2 P); 125.4 (qd, ¹ J_{CF} 275, ² J_{CP} 3.1, CF₃). ¹⁹F: -57.5 (td, ³ J_{FH} 11.75, ³ J_{FP} 8.7), -75.8 (s, TFA). ³¹P: 23.6 (broad s). MS-ESI: (+): 610.4 ([M + H]⁺, calc. 610.1); (-): 608.3 ([M - H]⁻, calc. 608.1), 722.3 ([M + TFA - H]⁻, calc. 722.1). TLC: iPrOH: conc. aq. NH₃:water 10:3:3; R_f 0.85 (5% aq. CuSO₄, blue spot). Purity was checked by HPLC (Fig. S1 and S3†) and ¹H, ¹³C, ¹⁹F and ³¹P NMR spectroscopy (Fig. S4–S7†).

The ligand H_3 notp^{tfe} for some experiments was further purified (removal of any remaining TFA) by chromatography on the strong cationic exchanger (Dowex 50, H^+ -form) with water

Dalton Transactions Paper

as an eluent; TFA is eluted off slightly earlier than H₃notp^{tfe} which is thus enriched in the later fractions. The fractions were analysed by ¹⁹F NMR spectroscopy and combined according to their purity. After repeated procedures, some amount of pure H₃notp^{tfe} was isolated as a colourless thick oil containing only some small amount of water as an impurity.

Preparation of solutions of the complexes for NMR studies

The ligand stock solution was prepared by dissolution of H_3 notp^{tfe} (600 mg, TFA-free) in water into a 25 ml volumetric flask. The exact ligand concentration (38.3 mM) was determined by ¹⁹F qNMR using standardized aq. solution of TFA similarly as described previously.⁴⁷

For determination of magnetic moments, the samples were prepared by mixing exactly the measured volume (200-300 µl) of the aq. ligand stock solution, the appropriate amount of the aq. stock solution of the metal ion salts (MnCl₂, (NH₄)₂Fe (SO₄)₂, CoCl₂, NiCl₂, CuCl₂; 0.95 equiv., concentration determined by chelatometry), appropriate amount of 1.50 M ag. NaOH (2.90 equiv.) and exactly measured volume (200-300 µl) of aq. HEPES buffer (0.1 M) with pH 7.4. The solutions of the Mn²⁺, Fe²⁺ and Cu²⁺ complexes were measured immediately (a fast complex formation) and the solutions of the Co²⁺ and Ni²⁺ complexes were equilibrated at 50 °C in a tightly closed vial overnight. To prepare the Cr³⁺ complex, the mixture of the weighed solid CrCl₃·6H₂O and the ligand stock solution (1.05 equiv.) with pH adjusted to ca. 6 with diluted aq. NH3 was heated in a tightly closed vial at 90 °C for 5 d. The Fe³⁺ complex was prepared by mixing the stock solution of Fe(NO₃)₃ (concentration determined by iodometry) and the ligand stock solution (1.05 equiv.), pH adjustment to ca. 5 with diluted aq. NH₃ and heating at 90 °C in a tightly closed vial overnight. Before measurement, t-BuOH (10 µl) was added to 350 µl of the buffered (pH 7.4) complex solution into an NMR tube, an insert tube containing 0.1% t-BuOH in D₂O was inserted and the ¹H NMR spectra were acquired. In the case of the Fe²⁺ complex, a fast formation of a colloidal precipitate was observed when handling the solution on air and, therefore, the complex was prepared under an Ar atmosphere, the solution was filtered through a microfilter and measured immediately.

Samples of the metal complexes of H_3 notp^{tfe} for T_1 and T_2 NMR experiments were prepared in 1 ml vials by mixing the ligand stock solution (500 μ l) with the stock solution (10 μ l) of the appropriate metal salt containing 0.9 equiv. of the metal ion (*i.e.* Mg(ClO₄)₂, CrCl₃, MnCl₂, FeCl₃, CoCl₂, NiCl₂, CuCl₂ and ZnCl₂). These stock solutions were prepared by dissolution of 9 equiv. of the respective metal salts in water (100 μ l). The pH of the mixtures was carefully adjusted by stepwise addition of diluted aq. NaOH to *ca.* 7 (in the case of the Fe³⁺ complex, diluted aq. NH₃ was used). Then, the vials were closed and the mixtures were heated overnight at 50 °C (in the case of the Cr³⁺ and Fe³⁺ complexes, the mixtures were heated at 90 °C for 5 d). After cooling, pH was adjusted to 7.4 by diluted aq. NH₃ (7.5 in the case of the Mg²⁺–H₃notp^{tfe} system).

Successful formation of the complexes was seen from the colour change (Cr^{3+} , Fe^{3+} , Co^{2+} , Ni^{2+} and Cu^{2+}) and was con-

firmed by NMR spectroscopy (Tables S1 and S2†) and mass spectrometry. The NMR and UV-Vis spectra of the complexes are shown in Fig. S8-S25.†

[Mg^{II}(notp^{tfe})]⁻: colourless. NMR (H₂O, pH 7.5): ¹⁹F: -57.2 (35%, broad q, ${}^{3}J_{\text{FH}} = {}^{3}J_{\text{FP}} \sim 10$); -57.0 (65%, free ligand, dt). MS-ESI (-): m/z 630.1 ([M]⁻, calc. 630.1).

[Cr^{III}(notp^{tfe})]: deep purple. NMR (H_2O , pH 7.4): ^{19}F : -46.0 (extremely broad). MS-ESI (+): m/z 659.1 ([M + H]⁺, calc. 659.0).

 $[Mn^{II}(notp^{tfe})]^-$: colourless. NMR (H₂O, pH 7.4): ¹⁹F: -41.8 (extremely broad). MS-ESI (-): m/z 661.2 ($[M]^-$, calc. 661.0).

[Fe^{III}(notp^{tfe})]: yellow. NMR (H₂O, pH 7.4): ¹⁹F: -31.0 (very broad). MS-ESI (+): m/z 663.1 ([M + H]⁺, calc. 663.0).

[Co^{II}(notp^{tfe})]⁻: purple-pink. NMR (D₂O, pD 7.4, evaporated to dryness and re-dissolved in D₂O): 1 H: -51.5, -12.8, 24.1, 83.4, 113.9, 115.2 and 157.0 (broad singlets). 13 C{ 1 H}: -461, -201, -115, 158 and 190 (all broad s). 19 F: -50.5 (broad). 31 P { 1 H}: 204 (broad). MS-ESI (+): m/z 667.2 ([M + 2H] $^{+}$, calc. 667.0).

[Ni^{II}(notp^{tfe})]⁻: light blue. NMR (H₂O, pH 7.4): ¹⁹F: -48.1 (91%, broad); -45.4 (9%, broad). MS-ESI (-): m/z 664.9 ([M]⁻, calc. 664.0).

 $[Cu^{II}(notp^{tfe})]^-$: deep blue. NMR (H₂O, pH 7.4): ¹⁹F: -54.2 (broad). MS-ESI (-): m/z 669.1 ([M]⁻, calc. 669.0).

[Zn^{II}(notp^{tfe})]⁻: colourless. NMR (D₂O, pD 7.4, evaporated to dryness and re-dissolved in D₂O): ¹H: 2.87 (broad s, 10H); 3.12 (broad s, 14H). ¹³C{¹H}: 36.61 (dq, ¹ $J_{\rm CP}$ 89.8, ² $J_{\rm CF}$ 28.5, PCH₂CF₃); 53.11 and 57.05 (2 × broad s, NCH₂CH₂N); 60.0 (d, ¹ $J_{\rm CP}$ 102, NCH₂P); 126.2 (q, ¹ $J_{\rm CF}$ 274, ² $J_{\rm CP}$ 1.5, CF₃). ¹⁹F: -57.21 (pseudo-q, ³ $J_{\rm FH}$ = ³ $J_{\rm FP}$ ~10.5). ³¹P: 29.02 (m, ³ $J_{\rm PH}$ = ³ $J_{\rm FF}$ ~10.5). ³¹P{¹H}: 29.02 (q, ³ $J_{\rm PF}$ 10.5). MS-ESI (+): m/z 670.1 ([M]⁻, calc. 670.0).

Electrochemical study

The stock solutions of the H_3 notp^{tfe} complexes for electrochemical experiments were prepared analogously as solutions for NMR studies but starting from the ligand stock solution (exactly 1.000 ml) and the corresponding solid metal salt (exactly 0.9 equiv.; $CrCl_3 \cdot 6H_2O$, $MnCl_2 \cdot 4H_2O$, $FeCl_3 \cdot 6H_2O$, $CoCl_2 \cdot 6H_2O$, $NiCl_2 \cdot 6H_2O$, $CuCl_2 \cdot 2H_2O$); The pH of the solutions was adjusted to 7.4.

The H_3 nota complexes for the electrochemical experiments with the same metal ions were prepared and isolated in the solid form as described in the literature (or by analogous procedures).⁵² The solids were dissolved in water before the measurement.

Cyclic voltammetry was performed using a potentiostat Polarographic analyzer PA 3 equipped with an XY writer (Laboratorní přístroje Praha). A three-electrode setup was used. Working electrodes were a hanging mercury drop electrode (HMDE) or platinum disc electrode; a saturated calomel electrode (SCE) was used as the reference electrode. Platinum wire with a plate was used as the auxiliary (counter) electrode. Among the several aqueous supporting electrolytes tested (0.1 M NaH₂PO₄/Na₂HPO₄ buffer with pH 7.4, 0.05 M LiClO₄, 0.05 M (NH₄)ClO₄, 0.05 M (NEt₄)ClO₄), 0.05 M aq. LiClO₄ was chosen due to the widest measurement window [+0.4–(-2.2) V

and +1.5-(-0.9) V for the HMDE and Pt electrode, respectively, vs. SCE]. For each measurement, 0.05 M aq. LiClO₄ (10 ml) was deoxygenated by bubbling argon through the solution in the measuring cell for several minutes; the absence of dissolved oxygen was confirmed by the measurement of a blank scan. Then, an appropriate volume of the stock solution of the

studied H₃notp^{tfe} complex or weighed amount of the solid H₃nota complex was added to reach a concentration of 1–4 mM for the complexes, and the solution was shortly deoxygenated again before the start of the electrochemical experiment.

Spectro-electrochemical experiments were performed using an optically transparent thin-layer electrochemical cell (OTTLE cell) assembled from a Pt mesh working electrode, Pt mesh auxiliary electrode and Ag wire as the reference electrode placed in a thin layer (optical path 0.2 mm) between two quartz windows. ⁵³ Electrodes were connected to a potentiostat WaveDriver 10 (Pine research) driven by Aftermath 1.6 software. The OTTLE cell was placed into a UV-Vis spectrophotometer UV-18010 (Shimadzu).

Electrosynthesis of the $[\text{Co}^{\text{II}}(\text{notp}^{\text{tfe}})]$ complex was performed in an H-shaped electrochemical cell with a fine frit placed in the middle of the horizontal part connecting two vertical tubes. One tube was equipped with the auxiliary electrode (Pt wire with a plate) and was filled with 0.05 M LiClO₄. The second tube contained the working electrode (Pt wire with a plate). The experiment was performed starting from 5 mM solution of the $[\text{Co}^{\text{II}}(\text{notp}^{\text{tfe}})]^-$ complex in 0.05 M LiClO₄.

Potentiometric study

The methodology of potentiometric titrations and processing of the experimental data were analogous to those previously reported. The ligand stock solution was prepared by dissolution of TFA-free H₃notp^{tfe} (600 mg) in a 50 ml volumetric flask. The exact ligand concentration (19.2 mM) was determined by TP qNMR using standardized aq. solution of TFA similarly as it was used previously. This value agreed well with the value calculated during the fitting of the protonation constants (19.38 mM, difference <1%) which was used for the calculations of the constants.

Potentiometric titrations were carried out in a vessel thermostatted to 25 \pm 0.1 °C. The titrations of the free ligand and systems containing Mg²⁺, Ca²⁺, Mn²⁺, Cu²⁺ and Zn²⁺ were performed in the -log[H⁺] range 1.6-12.1. The concentration of the ligand in the titration vessel was ca. 0.004 M, ligand-tometal ratio 1:0.95, ionic strength 0.1 M (NMe₄)Cl and starting volume ca. 5 ml. An equilibrium in the systems Co²⁺-H₃notp^{tfe} and Ni²⁺-H₃notp^{tfe} was established slowly and, therefore, the out-of-cell method was used in these cases. Each solution corresponding to one titration point was prepared in a tube with a ground joint (pH 1.6-6.5, three titrations with 15 points, starting volume ca. 1 ml, the same concentrations as used in a common titration) and the solutions were left to equilibrate at room temperature for one week. The overall protonation constants β_h are concentration constants and are defined by $\beta_h = [H_h L]/([H]^h \cdot [L])$ (stepwise protonation constants

are defined as $\log K_1 = \log \beta_1$; $\log K_h = \log \beta_h - \log \beta_{h-1}$). The overall stability constants are defined by the general equation $\beta_{\rm hml} = [\rm M_m H_h L_l]/([\rm M]^m \cdot [\rm H]^h \cdot [\rm L]^l)$. Here, the formation of only M:L 1:1 complexes (m=l=1) was considered. The water ion product, pK_w 13.81, and stability constants of $\rm M^{2^+} - \rm OH^-$ systems were taken from the literature. ⁵⁵ The constants (with their standard deviations) were calculated with program package OPIUM (Table S3†). ⁵⁶

Single-crystal X-ray diffraction study

Preparation of single crystals. The single crystals of $(NH_4)_3[Mn(notp^{tfe})]Cl_2\cdot 3H_2O$ and $(NH_4)_2[Cu(notp^{tfe})]Cl\cdot 3H_2O$ were prepared by neutralization of an aq. solution containing equimolar amounts of the ligand and MCl₂ (M = Mn, Cu) with diluted aq. NH₃ to pH ca. 7 followed by a slow evaporation. Single crystals of [Mn(H₂O)₆][Mn(notp^{tfe})]₂·18H₂O were obtained by a similar procedure but an excess of MnCl2 was used and some Mn(OH)2 was filtered off before the solvent evaporation. The crystals of (NH₄)[Co(notp^{tfe})]·3.5H₂O, (NH₄) [Cu(notp^{tfe})]·3.5H₂O and [Mg(H₂O)₆][Ni(notp^{tfe})]₂·12H₂O were prepared from the corresponding aq. solutions of the (NH₄)[M (notp^{tfe})] complexes by the slow vapour diffusion of EtOH. The ammonium salts (NH₄)[M(notp^{tfe})] (M = Co, Cu) were obtained by reaction of the ligand with a slight excess of freshly precipitated M(OH)₂ followed by chromatography of the formed complex on silica using aq. NH3:EtOH in a 1:10 mixture as an eluent; under these conditions, the excess of free M²⁺ was kept at the top of the column. In the case of [Mg(H2O)6][Ni (notp^{tfe})]₂·12H₂O, the ammonium salt obtained by chromatography was crystallized by concentration of the aq. solution in the presence of an excess of MgCl2 (10 equiv.). The single crystals of [Co(H₂O)₆][Co(notp^{tfe})]₂·14.25H₂O·0.75MeOH were prepared by saturation of the ligand solution with an excess of freshly precipitated Co(OH)2, filtration, concentration and a slow vapour diffusion of MeOH into the complex solution. The single crystals of Na₃[Co(notp^{tfe})]₂Br·3Me₂CO were prepared by neutralization of an equimolar aq. solution of the ligand and CoBr₂ with NaOH to pH ca. 7, concentration and a slow vapour diffusion of acetone. The single crystals of [ZnCl(H2O)3] [Zn(notp^{tfe})]-2H₂O were obtained by mixing the ligand and an excess of ZnCl2, neutralization with diluted aq. NH3 to pH ca. 5. and a slow concentration in air.

General procedure for data acquisition and treatment. The diffraction data were collected using a Bruker D8 VENTURE Duo diffractometer with an I μ S micro-focus sealed tube using Cu-K $_{\alpha}$ radiation (λ 1.54178 Å) or Mo-K $_{\alpha}$ radiation (λ 0.71073 Å). Data were analysed using the SAINT software package (Bruker AXS Inc., 2015–2019). Data were corrected for absorption effects using the multi-scan method (SADABS). All structures were solved by direct methods (SHELXT2014) and refined using full-matrix least-squares techniques (SHELXL2017). The details of the structure refinements are given in the ESI. For overview of the experimental crystallographic data, see Table S4.† The cif-files of all structures reported here have been deposited in the Cambridge Crystallographic Data Centre (CCDC 2327147–2327158†).

Dalton Transactions Paper

Results and discussion

Synthesis of the ligand H₃notp^{tfe}

Synthesis of the studied ligand H₃notp^{tfe} (H₃L) was performed as shown in Scheme 1.

The synthesis of the starting (2,2,2-trifluoroethyl)phosphinic acid was reported previously;⁴⁷ however, the use of commercial 2,2,2-trifluoroethyliodide for an Arbuzov-type reaction with bis(trimethylsilyl)hypophosphite resulted in the presence of hydroiodic acid in the final mixture that complicated further purifications.⁴⁷ Therefore, we optimized the synthesis using 2,2,2-trifluoroethylbromide. This compound was prepared by reaction of 2,2,2-trifluoroethyl-tosylate⁴⁹ with KBr in boiling anhydrous diethyleneglycol (boiling point 244 °C).⁵⁰ However, the synthesis needs non-trivial apparatus as the product is volatile (boiling point 26 °C) (see Experimental and Fig. S2†).

A Mannich-type reaction between tacn, (2,2,2-trifluoroethyl) phosphinic acid and paraformaldehyde was performed in a strong acid solution (50% v/v aq. TFA) at 40 °C. It was found that only a slight excess of (2,2,2-trifluoroethyl)phosphinic acid (3.1 equiv.) and paraformaldehyde (3.5 equiv.) is sufficient to reach almost quantitative conversion according to the ¹⁹F and ³¹P NMR spectra. Only a small extent of oxidation of (2,2,2-trifluoroethyl)phosphinic acid to (2,2,2-trifluoroethyl) phosphonic acid and formation of hydroxymethyl-(2,2,2-trifluoroethyl)phosphinic acid were observed under these conditions. The ligand H₃notp^{tfe} behaves as a strong acid and can be eluted from a strong cation exchange resin using water. However, its elution is only slightly slowed down compared to the simple acids present in the mixture [TFA, (2,2,2-trifluoroethyl)phosphonic acid, hydroxymethyl-(2,2,2-trifluoroethyl) phosphinic acid, an excess of starting (2,2,2-trifluoroethyl) phosphinic acid]. Traces of macrocyclic by-products with two phosphinic acid pendant arms (a twice-substituted intermediate and its N-methylated derivative, Scheme 1) were kept on the resin and could be eluted using 5% aq. ammonia solution. The ligand H₃notp^{tfe} was further purified by repeated chromatography on silica with conc. aq. NH3: EtOH 1:25 as the mobile phase to get a non-stoichiometric ammonium salt or by flash chromatography on a reverse phase column using a water: acetonitrile gradient containing 0.1% TFA as a modifier

Scheme 1 Synthesis of the studied ligand H₃notp^{tfe}.

to obtain non-stoichiometric trifluoroacetate. The ammonium salt of the ligand can be converted to the free ligand H₃notp^{tfe} on a strong cation exchange resin; ammonium cations stay on the column and the acidic form of H₃notp^{tfe} is eluted with water. In the case of the H₃notp^{tfe}–TFA adduct, a similar procedure can be used; however here, TFA is eluted only slightly earlier than the free H₃notp^{tfe} ligand and, thus, the procedure must be repeated to remove TFA completely. The ligand was isolated as a hygroscopic thick semi-solid glassy material, which complicates further analytical work. Therefore, concentrations of its stock solutions had to be determined independently by ¹⁹F qNMR using the addition of a known amount of standardized TFA solution.

Equilibrium studies

To study the acid-base behaviour of H₃notp^{tfe} and the stability of its complexes, potentiometric titrations were employed. Two protonation constants were calculated from the titration data acquired in the pH range 1.6-12.1. Tables 1 and S3[†] list the results together with data reported for related ligands. The observed protonation steps correspond to the protonations of the amino groups of the tacn skeleton. Going from the fully deprotonated species, the first protonation proceeds in the alkaline region ($\log K_1 > 10$) as it is common for other tacnbased ligands; the observed value falls into a range of values for other tacn-tris(phosphinic acid) derivatives (Table 1).60-62 Compared to the acetate (H₃nota) and phosphonate (H₆notp) derivatives (Fig. 1), the basicity of H₃notp^{tfe} is lower due to the electron-withdrawing characteristic of the phosphinate groups. 63 The protonation constant describing the second protonation step also corresponds well to those of other tacn-tris (phosphinic acid) derivatives (Table 1). Further protonations lay below the pH range accessible by potentiometry pointing to the strong acidity of the phosphinic acid groups. The calculated distribution diagram of differently protonated H3notptfe species is shown in Fig. S26.†

The thermodynamic stabilities of the complexes with selected alkaline earth (Mg²⁺, Ca²⁺) and transition (Mn²⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺) metal ions were studied in the M²⁺-H₃notp^{tfe} systems using a slight ligand excess. Except for Co²⁺ and Ni²⁺, complexation of all metal ions was fast enough for conventional potentiometry. The systems containing Co²⁺/Ni²⁺ ions were studied by the out-of-cell method. The system containing Fe2+ cannot be studied by this method as some precipitate is irreversibly formed during the titration. In general, besides formation of the [M(notp^{tfe})] complexes, the formation of hydroxido-complexes [M(notp^{tfe})(OH)]²⁻ was observed in strongly alkaline solutions pointing to the weak nucleophilicity of the pendant arm oxygen atoms which are replaced by the hydroxido ligand in a strongly alkaline solution (Table 1). It should be noted that potential formation of the hydroxido complexes of Co2+ and Ni2+ ions cannot be confirmed as the out-of-cell titration used to study the systems cannot be reliably performed up to the alkaline region. Stability constants of [M (notp^{tfe})] complexes are similar to those found for other phosphinic acid derivatives and are significantly lower in compari-

Table 1 Logarithms of ligand protonation constants ($\log K_h$), stability constants of the studied metal complexes ($\log \beta_{011}$) and protonation constants of hydroxidocomplexes ($\log \beta_{011} - \log \beta_{-111}$) of H₃notp^{tfe}, and comparison with related ligands. Charges of the species are omitted for clarity; "L" means fully deprotonated form of the ligand

Equilibrium	H ₃ notp ^{tfe a}	H_3 not p^H	H_3 not p^{Me}	H ₃ nota	H ₆ notp
$\mathbf{H}^+ + \mathbf{L} = \mathbf{H}\mathbf{L}$	10.23	10.16 ^b	10.92^{d}	13.17 ^e	12.1 ^g
		10.48^{c}			11.9 ^h
		,	7		11.79 ⁱ
$H^+ + HL = H_2L$	2.86	3.13 ^b	3.97^{d}	5.74^{e}	9.4_{b}^{g}
		3.28^{c}			9.3^{h}_{i}
+		h	d	6	8.65^{i}
$H^+ + H_2L = H_3L$	_	1.11^{b}	2.09^{d}	3.22^e	7.5 ^g
					7.6^{h}
$H^+ + H_3L = H_4L$				1.96^e	7.09 ^{<i>i</i>} 5.9 ^g
$\Pi + \Pi_3 L - \Pi_4 L$	_	_	_	1.90	5.4^{h}
					5.38^{i}
$H^+ + H_4L = H_5L$	_	_	_	0.70^{f}	2.9^g
43-				****	2.7^{h}
					2.53^{i}
$Mg^{2+} + L = [Mg(L)]$	5.04	5.32^{b}	6.66^{d}	10.97 ^c	11.01^{i}
$[Mg(L)(OH)] + H^{+} = [Mg(L)]$ $Ca^{2+} + L = [Ca(L)]$	11.71	– .	- .	_	— .
$Ca^{2+} + L = [Ca(L)]$	3.83	4.29^{b}	4.45^{d}	10.32^{c}	6.38 ⁱ
$\left[\operatorname{Ca}(L)(\operatorname{OH})\right] + \operatorname{H}^{+} = \left[\operatorname{Ca}(L)\right]$	12.87	11.70^{b}	_	_	
$Mn^{2+} + L = [Mn(L)]$	10.61	_	_	16.30^{e}	16.6 ⁱ
$[Mn(L)(OH)] + H^{+} = [Mn(L)]$	10.17		_		
$Co^{2^{+}} + L = [Co(L)]$ $Ni^{2^{+}} + L = [Ni(L)]$	13.04	12.97^{b}	_	20.13^{f}	19.7^{i}
$Ni^{2^{+}} + L = [Ni(L)]$ $Cu^{2^{+}} + L = [Cu(L)]$	13.18	$13.40^{b} \\ 13.43^{b}$	_	$19.24^{f} \ 23.33^{f}$	$19.4^{i} \ 21.3^{i}$
$Cu + L = [Cu(L)]$ $[Cu(I)(OH)] + H^{+} = [Cu(I)]$	13.50	13.43	_	23.33	21.3
$[Cu(L)(OH)] + H^{+} = [Cu(L)]$ $Zn^{2^{+}} + L = [Zn(L)]$	11.57 13.40	$\frac{-}{13.04^{b}}$	_	$\frac{-}{22.32^f}$	$\frac{-}{24.9^i}$
$\left[\operatorname{Zn}(L)(\operatorname{OH})\right] + \operatorname{H}^{+} = \left[\operatorname{Zn}(L)\right]$	12.26	13.04	_		24.3 —

^a This work, 0.1 M (NMe₄)Cl. ^b 0.1 M KNO₃, ref. 60. ^c 0.1 M (NMe₄)Cl, ref. 61. ^d 0.1 M KCl, ref. 62. ^e 0.1 M (NMe₄)Cl, ref. 64. ^f 0.1 M (NMe₄)Cl, ref. 65. ^g 0.1 M (NMe₄)Cl, ref. 66. ^h 0.1 M KNO₃, ref. 67. ⁱ 1 M KNO₃, ref. 68.

son with those of H_3 nota and H_6 notp as a result of a generally lower orverall basicity of the donor sites in the phosphinate ligands. The ligand H_3 notp^{tfe} shows a slight selectivity for Mg^{2+} over Ca^{2+} analogously to other related derivatives; however, these biogenic ions are not fully complexed even in the strongly alkaline region (see Fig. 2 and S27†).

The complexes with the transition metal ions are much more stable than those of alkaline earth ions; the Mn^{2+} complex has the lowest stability among the studied transition metal ions and is fully formed above pH 7 (Fig. 2 and S28†). Complexes with Co^{2+} – Zn^{2+} have comparable stability and the ligand shows almost no selectivity for the Cu^{2+} ion; so, the effect of Williams–Irving ordering is negligible. These metal ions are fully complexed at pH > 3; an illustrative distribution diagram is shown in Fig. 2 and S29.†

The observed non-selectivity for the Cu^{2+} ion is consistent with the non-selectivity of other phosphorus-containing tachbased ligands but it is in contrast to the selectivity observed for H_3 nota (Table 1). Overall, the ligand H_3 notp^{tfe} binds paramagnetic transition metal ions into stable complexes with a high selectivity over biogenic alkaline earths ions which is promising for potential *in vitro/in vivo* use.

Magnetic properties of the complexes in solution

As the value of magnetic moment plays a significant role in the paramagnetic relaxation enhancement mechanism, we

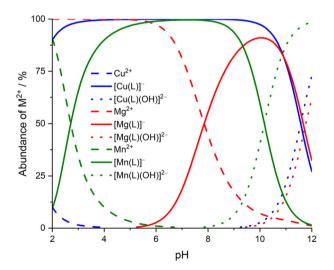


Fig. 2 Distribution diagrams of the $M^{2+}-H_3notp^{tfe}$ (= $M^{2+}-H_3L$) systems; $c(M^{2+})=c(H_3notp^{tfe})=0.004$ M, M = Mg, Mn and Cu. The abundances of the species formed in the $Mg^{2+}-H_3notp^{tfe}$ system are shown in red, those present in the $Mn^{2+}-H_3notp^{tfe}$ system are shown in green and those present in the $Cu^{2+}-H_3notp^{tfe}$ system are shown in blue. Dashed lines represent abundances of the free M^{2+} ions, full lines represent the $[M^{II}(notp^{tfe})]^-$ species, and the dotted lines represent the $[M^{II}(notp^{tfe})]^-$ species.

Dalton Transactions

determined the magnetic moments of the studied paramagnetic complexes in solution. As the Mn²⁺/Cu²⁺-H₃notp^{tfe} complexes are formed quickly (see Potentiometric study section), they were prepared by mixing the metal salt and ligand (in a slight excess) stock solutions, and adjusting the pH to 7.4. Solutions of the Co²⁺ and Ni²⁺ complexes were prepared analogously but the solutions were heated to 50 °C overnight to ensure full complexation. In the system containing Fe²⁺, some colloidal precipitate was continuously formed; therefore, measurement of the magnetic moment was performed immediately after preparation of the solution under argon and filtration through a microfilter.

Solutions of the Cr³⁺ and Fe³⁺ complexes were prepared by the reaction of CrCl₃·6H₂O (solid) or Fe(NO₃)₃ (stock solution), respectively, with a slight excess of H₃notp^{tfe} after neutralization to pH 5-6 and heating at 90 °C for 5 d (Cr3+) or overnight (Fe³⁺). Surprisingly, attempts to prepare a solution of the Co³⁺ complex by reaction of the ligand with Na₃[Co(CO₃)₃] (a kinetically labile precursor commonly used for a direct preparation of trivalent cobalt complexes)⁶⁹ failed; the Co²⁺ complex was formed instead as identified by UV-Vis and 19F NMR spectroscopies.

Magnetic moments were determined using Evans' method^{70,71} from the chemical shift difference of t-BuOH present in the solution of the complex with a known concentration and aq. solution in the insert cuvette and are listed in

The values of $\mu_{\rm eff}$ clearly show that the complexes of ${\rm Mn}^{2+}$, Fe²⁺/Fe³⁺ and Co²⁺ are high-spin and, thus, an overall ligand field induced by the (notp^{tfe})³⁻ anion is relatively small.

Redox properties of the complexes

The redox behaviour of transition metal complexes of the "parent" H₃nota is well documented in the literature. ^{52,72} Redox potentials of the M^{3+/2+}-H₃nota systems are reported for M = Cr, Mn, Fe, Co and Ni, and values of some of them fall into a biologically relevant range (Table 3). The change of oxidation state governs electronic and magnetic properties of the ions, and can significantly alter the chemical shift of the 19F NMR signal and its relaxation properties. If the redox change can occur under in vivo conditions, the complex can be utilized as a redox-responsive probe. 18 Therefore, electrochemical studies of H₃notp^{tfe} complexes and analogous H₃nota complexes were performed to obtain directly comparable data as

Table 2 Magnetic moments μ_{eff} of the $M^{2+}/M^{3+}-H_3$ notp^{tfe} complexes in aq. solution (pH 7.4)

Complex	$\mu_{\mathrm{eff}}\left(\mathrm{B.M.}\right)$
[Cr ^{III} (notp ^{tfe})]	3.51
[Mn ^{II} (notp ^{tfé})] ⁻	5.75
Fe ^{II} (notp ^{tfe})	5.29
[Fe ^{III} (notp ^{tfé})]	6.16
[Co ^{II} (notp ^{tfe})] ⁻	4.98
[Ni ^{II} (notp ^{tfe})] ⁻	2.77
[Cu ^{II} (notp ^{tfé})]	1.86

Table 3 Redox potentials of $[M^{III}(L)]/[M^{II}(L)]^-$ pairs of the transition metal ion complexes of $L = H_3 notp^{tfe}$ and $H_3 nota$, obtained by CV (0.05 M aq. LiClO₄, ambient temperature, values are given in V vs. SCE)

	M. IX as to the	M-H ₃ nota	
Redox pair	M-H ₃ notp ^{tfe} This work	This work	Literature ⁵²
Cr ³⁺ /Cr ²⁺	-1.32^{a}	-1.39^{a}	-1.41
Mn^{3+}/Mn^{2+}	0.67^{b}	0.56^{b}	0.56
Mn^{4+}/Mn^{3+}	0.96^{b}	1.02^{b}	_
Fe ³⁺ /Fe ²⁺	$-0.02^{a,b}$	-0.035^{a}	-0.045^{b}
		-0.045	
Co ³⁺ /Co ²⁺	0.95^{b}	-0.23^{a}	-0.24
Ni ³⁺ /Ni ²⁺	<u></u> c	0.95^{b}	0.92

^a Hanging mercury drop electrode. ^b Platinum electrode. ^c No electrochemical process was observed.

data reported in the literature were sometimes acquired under different or unspecified conditions (electrode material, supporting electrolyte). Besides the metal ions mentioned above, we also studied the Cu²⁺ complex as the Cu²⁺/Cu⁺ pair can also be employed as a redox probe. 18

0.05 M aq. LiClO₄ solution was chosen as the supporting electrolyte due to a wide measurement window [+0.4-(-2.2) and 1.5-(-0.9) V using the HMDE and Pt electrode, respectively, vs. SCE], and the complexes were studied by cyclic voltammetry (CV). The reversible behaviour of the M²⁺/M³⁺- H_3 notp^{tfe} and M^{2+}/M^{3+} - H_3 nota pairs was found for M = Cr, Mn, Fe and Co (Table 3). In contrast to the [Ni^{II}(nota)] complex, [Ni^{II}(notp^{tfe})]⁻ cannot be oxidized to the [Ni^{III}(notp^{tfe})] complex in the accessible potential range.

In the case of the Mn-systems with both ligands, a further reversible process was observed which is attributable to the [Mn^{IV}(L)]⁺/[Mn^{III}(L)] redox pair. This behaviour was reported for the Mn²⁺-H₃nota complex previously⁷² but the reported potentials of 0.30 and 0.68 V for the [Mn^{III}(nota)]/[Mn^{II}(nota)] and [Mn^{IV}(nota)]⁺/[Mn^{III}(nota)] pairs, respectively, are not consistent with other literature data⁵² and with our value; probably, the reported potentials were wrongly corrected for the SCE potential. To identify the species and to study their stabilities, a spectro-electrochemical study was performed. If the potential was increased in the range from 0.5 to 1.0 V, a d-d transition band centred at 470 nm gradually appeared (Fig. S30†), attributable to Mn³⁺ due to the one-electron oxidation of [Mn^{II}(L)]⁻ and the formation of [Mn^{III}(L)], consistent with the results obtained by the CV study. Further oxidation (applied potential up to 1.8 V) led to a gradual increase in absorbance over the whole spectral range, with no distinguished absorption band (Fi S31†). It was probably caused by the formation of a colloidal precipitate. Thus, one can conclude that the oxidation of [Mn^{III}(L)] to [Mn^{IV}(L)]⁺ is pseudoreversible on the short CV timescale but the formed Mn^{IV}species decomposes and colloidal MnO2·nH2O is formed during the longer spectro-electrochemical timescale.

The spectro-electrochemical reduction (applied potential from 0.5 to -0.8 V) of the [Fe^{III}(notp^{tfe})] complex led to the formation of [Fe^{II}(notp^{tfe})]⁻ as documented by the gradual spec-

tral change (Fig. S32†). The [Fe^{II}(notp^{tfe})]⁻ species was found

to be stable on the given timescale (order of seconds); however as observed earlier, it decomposes on longer standing (see the Magnetic properties section).

The electro-synthetic oxidation (applied potential 1.1 V) of the [Co^{II}(notp^{tfe})]⁻ complex (characterized by three close lowintensity absorption bands centred around 520 nm) revealed the formation of the [Co^{III}(notp^{tfe})] complex (the appearance of two new intense d-d bands at 394 and 557 nm, as shown in Fig. S33†) which is stable in solution for at least several days.

The Cu²⁺ complexes of both H₃notp^{tfe} and H₃nota underwent irreversible two-electron reduction on HMDE as proved by the anodic peak corresponding to the oxidation of metallic copper; no electrochemical changes were observed using the Pt electrode. According to the measured values, the [Cr^{II}(notp^{tfe})] complex is a very strong reduction agent similar to [CrII(nota)]-, and its potential lies close to the reduction edge of HDME; such a value cannot be practically utilized in biological systems. However, the potentials of [M^{III}(L)]/ $[M^{II}(L)]^{-}$ pairs with M = Mn, Fe and Co lie in the biologically relevant range. Among them, the stability of the Mn²⁺ complex is not sufficient for utilization in biological systems (full complexation at pH > 7, see Potentiometric study section) and the [Fe^{II}(notp^{tfe})] species was found to be kinetically labile, so only the Co³⁺/Co²⁺-H₃notp^{tfe} redox pair is suitable for *in vitro*/ in vivo utilization. Furthermore, the [Co^{II}(notp^{tfe})] complex is a high-spin paramagnetic species but the oxidized form [Co^{III}(notp^{tfe})] is a low-spin diamagnetic complex, which induces a very significant change in the ¹⁹F NMR spectra (see below). Therefore, we studied the corresponding redox process using chemical oxidizing/reducing agents. The [Co^{III}(notp^{tfe})] species can be prepared by chemical oxidation of [Co^{II}(notp^{tfe})] with $K_2S_2O_8$ or H_2O_2 (Fig. S33†). The [Co^{III}(notp^{tfe})] species can be reduced back to the [Co^{II}(notp^{tfe})] complex using NaBH₄; other reducing agents tested - N2H4, NH2OH, Na2S2O4 - caused no reaction. It should be further highlighted that the reaction of H₃notp^{tfe} with Na₃[Co^{III}(CO₃)₃] afforded [Co^{II}(notp^{tfe})]⁻ and not the expected [CoIII(notptfe)] complex. Therefore, although the $[Co^{III}(notp^{tfe})]$ species is a relatively strong oxidation agent $(E_{\underline{1}})$ + 0.95 V vs. SCE, Table 3), some kinetic barrier plays a role in the tested chemical reductions, probably due to a full wrapping of the central metal ion with the ligand which prevents a close contact with a reduction agent. A comparison of the scaled UV-Vis spectra of [Co^{III}(notp^{tfe})] and [Co^{II}(notp^{tfe})] is shown in Fig. 3.

Solid-state structures of the complexes

The H₃nota-like ligands usually wrap the metal ions in a hexadentate fashion. Three nitrogen atoms of the tacn skeleton form a basal N₃-plane making one face of the coordination polyhedron, and the oxygen atoms of three pendant arms form a parallel upper O₃-plane. Formation of the chelate rings upon a coordination of the tacn unit results in a clock-wise or anticlock-wise torsion on the ethylene groups connecting the coordinated nitrogen atoms (i.e., in the Newman projection of

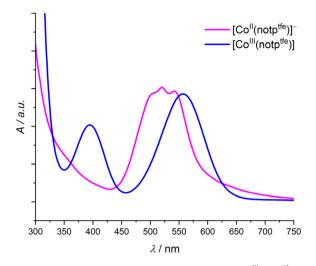


Fig. 3 Comparison of the UV-Vis spectra of $[Co^{III}(notp^{tfe})]$ and [Co^{II}(notp^{tfe})] complexes. Spectra are arbitrarily scaled. Extinction coefficients in dm³ mol⁻¹ cm⁻¹: $[Co^{II}(notp^{tfe})]^{-}$: $\varepsilon(502 \text{ nm})$ 16, $\varepsilon(520 \text{ nm})$ 17, ε (541 nm) 16; [Co^{III}(notp^{tfe})]: ε (394 nm) 68, ε (557 nm) 92.

the formal ethylene-diamine fragment, it depends on mutual positions of the C-N bonds of the front and rear carbon atoms, respectively); such conformations are denoted as δ or λ . All three chelate rings formed in the macrocyclic part have to adopt the same geometry due to steric hindrances, i.e. one of two possible macrocycle conformations δδδ or λλλ is formed. The pendant arm coordination leads to a torsion of the pendant arm with respect to the (pseudo)trigonal axis. Here, stereodescriptors Δ and Λ are used for clockwise and anticlockwise arrangements, respectively [i.e., differing in mutual positions of the O and N atoms belonging to the individual pendant arms in a view along the (pseudo)trigonal axis from the top to the bottom]. If the complex molecule has (pseudo) trigonal symmetry, all pendants are twisted in the same direction. A combination of both rotations leads to the formation of two diastereomeric pairs of enantiomers, $\Delta\Delta\Delta\lambda\lambda\lambda$ + $\Lambda\Lambda\Lambda\delta\delta\delta$ and $\Delta\Delta\Delta\delta\delta\delta + \Lambda\Lambda\Lambda\lambda\lambda\lambda$ (hereafter for simplicity denoted as $\Delta\lambda$ + $\Lambda\delta$ and $\Delta\delta$ + $\Lambda\lambda$). Analogous diastereoisomerism is welldocumented in the complexes of H₄dota-like ligands (Fig. 1) where it leads to the square-antiprismatic (SA isomer, $\Delta\lambda + \Lambda\delta$, real pendant torsion >35°) and the twisted-square-antiprismatic (TSA isomer, Δδ + Λλ, real pendant torsion <30°) geometries. 63,73-80 Similarly to the complexes of H₄dota-like ligands, the diastereoisomers of the complexes of H₃nota-like ligands also differ in a mutual twist of the N₃- and O₃-planes. The larger twist found in the $\Delta\lambda + \Lambda\delta$ isomer leads to a shorter separation of the N₃- and O₃-planes and to a geometry closer to an octahedron (OC). The $\Delta\delta+\Lambda\lambda$ conformations lead to the twisted-trigonal-antiprismatic (TTA) arrangement with a somewhat larger distance between the N₃- and O₃-planes. In the case of the H₃notp^{tfe} ligand, a further source of isomerism comes also from the absolute (R/S) configuration of the phosphorus atom upon coordination of the phosphinate group. Three pendant arms with potentially independent

Dalton Transactions Paper

R/S-isomerism give rise to a number of isomers (Fig. 4), similarly as observed for the phosphinate analogues of H₄dota. 81-83

Two Mn²⁺ phases were obtained where the central Mn²⁺ ion has the TTA coordination sphere. The absolute configuration on a phosphorus atom of all coordinated pendant phosphinate groups in the complex molecule is the same (in the latter compound forced due to the crystallographic trigonal symmetry of the space group, $R\bar{3}$), resulting in the presence of the enantio-

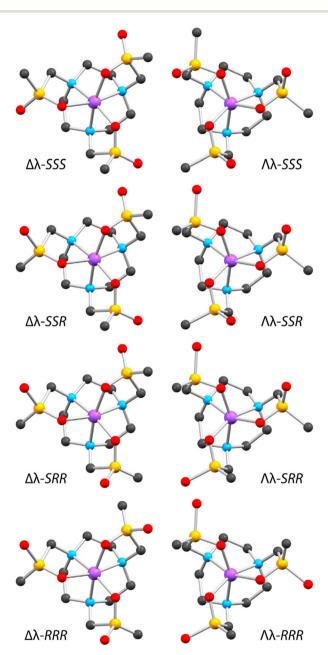


Fig. 4 Possible diastereoisomers of H_3 notp^{tfe} complexes; only one of the enantiomers of each diastereoisomer is shown. All shown enantiomers have the $\lambda\lambda\lambda$ conformations of the macrocycle, the other enantiomers have the $\delta\delta\delta$ conformations. Left column: the OC species; right column: the TTA species. Colour code: metal ion - magenta, phosphorus - orange, oxygen - red, nitrogen - light blue, carbon - dark grey.

meric pair $\Delta\delta$ -SSS and $\Lambda\lambda$ -RRR in both structures. Selected relevant geometric parameters are listed in Tables 4 and S5† and molecular structures of the complex species are shown in Fig. 5 and 6.

The coordination cages are relatively large compared to other complexes (see below) as evidenced by the distance between the N₃- and O₃-planes (2.9 and 2.8 Å, respectively). It is caused by the long length of the N-Mn coordination bonds. The large separation of the planes leads also to a relatively small twist angle between the planes (i.e. pendant arm rotation, ca. 8° and 24° for the complex anions in (NH₄)₃[Mn (notp^{tfe})]Cl₂·3H₂O $[Mn(H_2O)_6][Mn(notp^{tfe})]_2 \cdot 18H_2O$ and phases, respectively). In the crystal structure of [Mn(H₂O)₆][Mn (notp^{tfe})]₂·18H₂O, two [Mn(notp^{tfe})]⁻ anions are head-to-head connected to the central [Mn(H2O)6]2+ cation through hydrogen bonds (Fig. 6); a similar structural motif was also found in some other crystal structures (see below).

Several Co²⁺-containing phases were obtained and, surprisingly, different diastereoisomers of the [Co(notp^{tfe})] anion were found. In the crystal structure of (NH₄)[Co (notp^{tfe})]·3.5H₂O, the $\Delta\lambda$ -SSS and $\Lambda\delta$ -RRR enantiomers with the OC geometry were found (Fig. 7 and S37†). In the structure of $[Co(H_2O)_6][Co(notp^{tfe})]_2 \cdot 14.25H_2O \cdot 0.75MeOH$ the $\Delta \delta$ -SSS and Λλ-RRR enantiomeric pairs with TTA geometries were found; for an example, see Fig. 8. These complex anions are connected to the $\left[\text{Co}(\text{H}_2\text{O})_6\right]^{2^+}$ cation through the hydrogen bonds (Fig. S38†) similarly as it was observed in the crystal structure of [Mn(H₂O)₆][Mn(notp^{tfe})]₂·18H₂O discussed above, although the compounds are not isostructural. In contrast to the structures of the other Co²⁺ phases, the sodium salt Na₃[Co(notp^{tfe})]₂Br·3Me₂CO presents a TTA enantiomeric Δδ-RRR/Λλ-SSS pair (Fig. 8 and S39†). Selected relevant geometric parameters for the structures are listed in Tables 4 and

The Ni²⁺ complex was successfully crystallized as [Mg $(H_2O)_6$ [Ni(notp^{tfe})]₂·12H₂O in the space group $R\bar{3}$ and with the lattice parameters close to those of [Mn(H₂O)₆][Mn (notp^{tfe})]₂·18H₂O; however, the compounds are not isostructural. As in the $[Mn(H_2O)_6][Mn(notp^{tfe})]_2 \cdot 18H_2O$ and [Co $(H_2O)_6][Co(notp^{tfe})]_2\cdot 14.25H_2O\cdot 0.75MeOH$ phases, two [Ni (notp^{tfe})] anions are head-to-head connected to the central hexaaqua unit via hydrogen bonds (Fig. S40†). The [Ni $(\text{notp}^{\text{tfe}})^{-}$ species form an octahedral $\Lambda \delta$ -RRR/ $\Delta \lambda$ -SSS pair (Fig. 9). For geometric parameters, see Tables 4 and S5.†

In the case of the Cu²⁺ complex, two different phases were isolated in the solid state. The compound (NH₄)[Cu (notp^{tfe})]-3.5H₂O is isostructural with (NH₄)[Co (notp^{tfe})]-3.5H₂O and even an analogous disorder was found (see the ESI†). The $\Delta\lambda$ -SSS/ $\Lambda\delta$ -RRR enantiomers with the octahedral geometry were found (Fig. S41 and S42†). In the other phase of composition (NH₄)₂[Cu(notp^{tfe})]Cl·3H₂O, only two pendant arms are coordinated and they are turned in mutually opposite directions (Fig. 10). Centrosymmetry of the space group $(P\bar{1})$ leads formally to $\Delta \Lambda - \lambda \lambda \lambda - SR/\Lambda \Delta - \delta \delta \delta - RS$ enantiomers. Very similar pentacoordinated geometry was previously reported for the Cu²⁺ complexes of H₆notp and H₃notp^{Ph}

Selected geometric parameters found in the crystal structures of the studied complexes $[L^{3-} = (\text{notp}^{\text{tfe}})^{3-}]$ Table 4

	$(\mathrm{NH}_4)_3[\mathrm{Mn}(\mathrm{L})]$	$(\mathrm{NH}_4)_3[\mathrm{Mn}(\mathrm{L})]$ $[\mathrm{Mn}(\mathrm{H}_2\mathrm{O})_6]$	$(NH_4)[Co(L)]$		$[{ m Co}({ m H}_2{ m O})_6][{ m Co} \ [{ m L}]_2 \cdot 14.25 { m H}_2{ m O} \cdot 0.75 { m MeOH}$	МеОН		$Na_3[Co$ (L)] ₂ Br·3Me ₂ CO	Ae_2CO	$(NH_4)[Cu(L)]$	$\left(\mathrm{NH_4}\right)_2[\mathrm{Cu}(\mathbf{L})]$	$[\operatorname{Mg}(\operatorname{H}_2\operatorname{O})_6]$	$\left[\operatorname{ZnCl}(\mathbf{H}_2\mathbf{O})_3 \right]$
Geometry	Cl ₂ ·3H ₂ O TTA Δδ-SSS	$[MII(L)]_{2}^{2} \cdot 18H_{2}^{2}O$ TTA^{α} $\Delta \delta - SSS$	$3.3H_2O$ OC $\Delta\lambda$ -SSS	$_{\Lambda\lambda\text{-}RRR}^{\text{TTA}}$	TTA Δδ-SSS	${\rm TTA}^a \\ \Lambda \lambda\text{-}RRR$	TTA^a $\Delta\delta$ -SSS	TTA Aλ-SSS	TTA Λλ-SSS	э.эн ₂ О OC Лδ- <i>RRR</i>	Cι·3π ₂ O square pyramidal ΔΛ0-λλλ-SR0	$[N](L)_{12}$. 12 H_2 O G^a $\Lambda\delta$ -RRR	[211(L)]:2H2O OC ^a Aδ-RRR
Distances (Å)													
M-N1	2.360(1)	2.335(2)	2.160(2)	2.168(2)	2.168(2)	2.187(2)	2.170(2)	2.154(8)	2.149(8)	2.251(1)	2.040(2)	2.103(1)	2.191(3)
M-N4	2.377(1)	2.335(2)	2.144(2)	2.171(2)	2.178(2)	2.187(2)	2.170(2)	2.163(8)	2.156(8)	2.035(1)	2.021(2)	2.103(1)	2.191(3)
M-N7	2.341(1)	2.335(2)	2.166(2)	2.189(2)	2.184(2)	2.187(2)	2.170(2)	2.162(8)	2.141(8)	2.116(1)	2.254(2)	2.103(1)	2.191(3)
M-011	2.123(1)	2.113(2)	2.062(2)	2.081(1)	2.092(1)	2.084(1)	2.098(2)	2.108(6)	2.127(6)	1.989(1)	1.972(1)	2.059(1)	2.063(3)
M-021	2.144(1)	2.113(2)	2.060(2)	2.100(1)	2.098(1)	2.084(1)	2.098(2)	2.104(6)	2.133(6)	2.006(1)	1.940(1)	2.059(1)	2.063(3)
M-031	2.117(1)	2.113(2)	2.125(2)	2.072(1)	2.075(1)	2.084(1)	2.098(2)	2.093(7)	2.086(7)	2.360(1)	:	2.059(1)	2.063(3)
$M \cdots NQ^b$	1.662(1)	1.633(2)	1.398(2)	1.437(1)	1.436(1)	1.455(2)	1.435(2)	1.411(5)	1.396(5)	1.358(1)	1.315(1)	1.323(1)	1.436(1)
$\text{M} \cdots \text{OQ}^{b}$	1.242(1)	1.140(2)	1.091(1)	1.217(1)	1.209(1)	1.256(2)	1.202(2)	1.234(4)	1.223(4)	1.127(1)	:	1.123(1)	1.138(1)
$_{q}$ $00\cdots0$ 0 0	2.9042	2.7726	2.4912	2.6529	2.6443	2.7110	2.6369	2.6445	2.6189	2.4991	ı	2.4458	2.5739
$\mathbf{M} {\cdots} \mathbf{F} 1^c$	6.005(1)	5.869(2)	5.640(2)	5.545(1)	5.540(2)	5.644(1)	5.544(2)	5.893(6)	5.887(7)	5.634(1)	5.697(1)	5.574(1)	5.662(3)
$\mathbf{M} \cdots \mathbf{F} 2^c$	6.415(1)	6.388(2)	6.275(2)	6.277(1)	6.310(2)	6.341(1)	6.294(2)	6.222(6)	6.242(6)	6.248(1)	6.168(1)	6.233(1)	6.309(3)
$\mathbf{M} \cdots \mathbf{F} 3^c$	6.877(1)	6.649(2)	6.583(2)	6.339(1)	6.380(1)	6.470(1)	6.406(2)	6.803(6)	6.823(6)	6.545(1)	6.582(1)	6.414(1)	6.556(3)
$\mathbf{M} {\cdots} \mathbf{F} 4^c$	5.958(1)	5.869(2)	5.651(2)	5.613(1)	5.514(2)	5.644(1)	5.544(2)	5.391(8)	4.559(7)	5.556(1)	5.505(1)	5.574(1)	5.662(3)
$\mathrm{M}{\cdots}\mathrm{F5}^c$	6.468(1)	6.388(2)	6.333(2)	6.284(1)	6.206(2)	6.341(1)	6.294(2)	5.910(8)	5.097(7)	6.271(1)	6.125(1)	6.233(1)	6.309(3)
$\mathbf{M} {\cdots} \mathbf{F} 6^c$	6.831(1)	6.649(2)	6.466(2)	6.428(1)	6.256(2)	6.470(1)	6.406(2)	6.637(8)	6.290(7)	6.409(1)	6.278(1)	6.414(1)	6.556(3)
$\mathbf{M} \cdots \mathbf{F} 7^c$	5.880(1)	5.869(2)	$5.005(3)^d$	5.638(1)	5.654(1)	5.644(1)	5.544(2)	5.765(6)	5.744(7)	5.237(2)	5.264(1)	5.574(1)	5.662(3)
$\mathbf{M} \cdots \mathbf{F} 8^c$	6.347(1)	6.388(2)	$5.956(3)^d$	6.305(1)	6.328(1)	6.341(1)	6.294(2)	6.346(6)	6.349(6)	6.094(2)	6.888(2)	6.233(1)	6.309(3)
$\mathbf{M} {\cdots} \mathbf{F} 9^c$	6.836(1)	6.649(2)	$6.455(3)^d$	6.438(1)	6.476(1)	6.470(1)	6.406(2)	(9)669.9	6.723(6)	6.664(2)	7.169(1)	6.414(1)	6.556(3)
$mean(M \cdots F)$	6.40	6.30	6.04	6.10				80.9		20.9	6.19	20.9	6.18
Dihedral angles $(^\circ)$	6												
$N1-NQ-OQ-O11^{b}$		24.38(9)	46.9(1)	33.43(8)	33.03(8)	29.08(8)	35.99(9)	34.7(4)	33.5(3)	44.82(6)	1	50.39(4)	43.0(2)
$N4-NQ-OQ-O21^{b}$		24.38(9)	49.6(1)	32.27(8)	33.45(8)	29.08(8)	35.99(9)	33.2(3)	33.2(3)	53.70(6)	1	50.39(4)	43.0(2)
$N7-NQ-OQ-O31^{b}$	8.79(6)	24.38(9)	46.0(1)	31.28(8)	32.00(8)	29.08(8)	35.99(9)	34.2(4)	34.7(4)	45.56(6)	I	50.39(4)	43.0(2)

^a The complex molecule possesses trigonal symmetry; F1 = F4 = F7, F2 = F5 = F8, F3 = F6 = F9. ^b NQ and OQ are centroids of the N_3 - and O_3 - planes, respectively. ^c Formal numbering: fluorine atoms F1-3 belong to the pendant arm attached to N_3 , fluorine atoms F4-6 to the pendant arm attached to N_3 , ^d More abundant position of the disordered trifluoroethyl group.

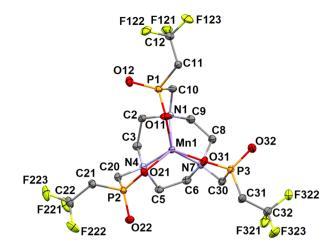


Fig. 5 Molecular structure of the [Mn(notp^{tfe})]⁻ anion with the $\Delta\delta$ -SSS TTA geometry found in the crystal structure of (NH₄)₃[Mn(notp^{tfe})] Cl₂·3H₂O. Hydrogen atoms are omitted for clarity.

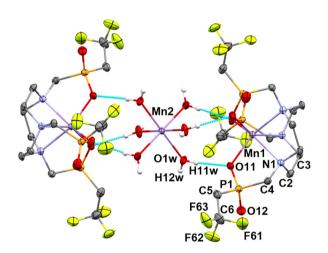


Fig. 6 Molecular structure of [Mn(H₂O)₆][Mn(notp^{tfe})]₂ fragment found in the crystal structure of [Mn(H₂O)₆][Mn(notp^{tfe})]₂·18H₂O. Intermolecular hydrogen bonds connecting two [Mn(notp^{tfe})]⁻ anions to the central [Mn(H₂O)₆]²⁺ cation are shown in turquoise. Individual [Mn (notp^{tfe})]⁻ species adopt the $\Delta\delta$ -SSS and $\Delta\lambda$ -RRR TTA geometries. Carbon-bound hydrogen atoms are omitted for clarity.

(Fig. 1). 84,85 Selected geometric parameters are listed in Tables 4 and S5. \dagger

Octahedral species with $\Lambda\delta$ -RRR and $\Delta\lambda$ -SSS geometry were also observed in the crystal structure of the Zn^{2+} complex, $[ZnCl(H_2O)_3][Zn(notp^{tfe})]\cdot 2H_2O$ (Fig. S43†). Here, the countercation $[ZnCl(H_2O)_3]^+$ is formed from the excess of $ZnCl_2$ used in the reaction. For geometric parameters, see Tables 4 and S5.†

Overall, the twisted trigonally antiprismatic (TTA, $\Delta\delta/\Lambda\lambda$) geometry is preferred for the largest ion (Mn²⁺) as this geometry leads to a larger coordination cage ($d_{\rm NQ\cdots OQ}$ 2.62–2.90 Å, Tables 4 and S5†). Smaller Ni²⁺, Cu²⁺ and Zn²⁺ ions form octahedral species (OC, $\Delta\lambda/\Lambda\delta$) with a shorter separation of the N₃-and O₃-planes (2.45–2.57 Å). The Co²⁺ ion of an intermediate

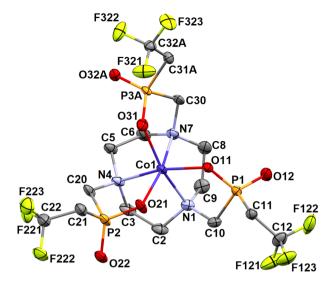


Fig. 7 Molecular structure of the $[Co(notp^{tfe})]^-$ anion with the $\Delta\lambda$ -SSS OC geometry found in the crystal structure of $(NH_4)[Co(notp^{tfe})]\cdot 3.5H_2O$. More abundant part of the disordered pendant arm (P3A and related atoms) is shown; for the disorder, see Fig. S37.† Hydrogen atoms are omitted for clarity.

size can adopt both environments, differing in this parameter. The torsion angles of the pendant arms are consistently smaller for the TTA (7–36°, ideal value 30°) than for the OC (43–54°, ideal value 60°) arrangements. Similar structural features were also found for divalent transition metal complexes of the related ligands. All reported divalent complexes of $\rm H_3notp^{\rm Ph}$ ($\rm Co^{2+}$, $\rm Ni^{2+}$, $\rm Cu^{2+}$ and $\rm Zn^{2+}$) adopt an OC geometry with the $\rm N_3$ –O₃ twist in the range 51–52°. ^{86,87} However, the Mn²⁺ complex of H₆notp also adopt the OC geometry but very distorted [twist 38°, distances from the centroids of the $\rm N_3$ -and O₃-planes, NQ and OQ, respectively, are $\it d(Mn\cdots NQ)$ 1.64 Å, $\it d(Mn\cdots NQ)$ 1.02 Å]. ⁸⁸

The critical parameter influencing the relaxation rates of the ¹⁹F nuclei is the distance between the paramagnetic ion and the fluorine atoms. Therefore, this parameter is compiled in Table 4 together with selected geometric parameters of the coordination cages. It can be seen that the exact geometry of the coordination sphere influences the M···F distance only negligibly – in the TTA species, the distances are only slightly longer (*ca.* 5.6–6.7 Å) when compared to the OC isomers (*ca.* 5.5–6.5 Å). Such distances fall in the range suggested to be relevant for a significant influence on the ¹⁹F NMR relaxation times. ⁸⁹ Very similar mean M···F lengths were observed also in the complexes of 1,8-H₂te2p^{tfe} (cyclam-based ligand with the same pendant arms, Fig. 1) which were studied previously. ⁴⁷

NMR spectral properties of the complexes

Measured NMR characteristics of the studied complexes are compiled in Tables 5, 6, S1 and S2.† A comparison of the 19 F NMR spectra of transition metal complexes of H_3 notp^{tfe} is shown in Fig. 11.

F3C **Ş**F3B C32A **03B** C31A **03B** C30A N7A C8A C₆A O₂A C₁₀A N1A C10A O2B C3A F₁B F2B 02B F₁B 01B

Fig. 8 Molecular structures of the $[Co(notp^{tfe})]^-$ anion with the $\Lambda\lambda$ -RRR TTA geometry found in the crystal structure of $[Co(H_2O)_6][Co(notp^{tfe})]_2$ 14.25H₂O·0.75MeOH (left) and that with the $\Lambda\lambda$ -SSS TTA geometry found in the crystal structure of Na₃[Co(notp^{tfe})]₂Br·3Me₂CO (right). Hydrogen atoms are omitted for clarity.

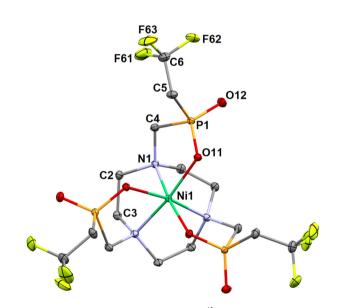


Fig. 9 Molecular structure of the [Ni(notp^{tfe})]⁻ anion with the $\Lambda\delta\text{-}RRR$ OC geometry found in the crystal structure of [Mg(H₂O)₆][Ni (notp^{tfe})]₂·12H₂O. Hydrogen atoms are omitted for clarity.

To study the solution structures of the complexes, the NMR spectra of diamagnetic (Mg^{2+}, Zn^{2+}) complexes also were acquired. The ^{19}F NMR spectrum of the Mg^{2+} – H_3 notp^{tfe} system at pH 7.5 fully agrees with the results of potentiometry – the spectrum revealed the presence of the well resolved signals of the free ligand and the complex in ca. 65:35 ratio (Fig. S8†). Besides these two species, some very minor signals were also observed, probably belonging to the complex species with

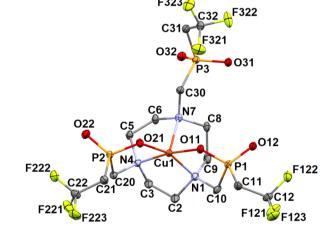


Fig. 10 Molecular structure of the pentacoordinated [Cu(notp^{tfe})]⁻ anion adopting the $\Delta\Lambda0-\lambda\lambda\lambda$ -SR geometry found in the crystal structure of (NH₄)₂[Cu(notp^{tfe})]Cl·3H₂O. Hydrogen atoms are omitted for clarity.

different R/S configurations of the phosphinate pendant groups or to some species in which some of the pendant arms are uncoordinated. It indicates that the ligand and complex species are not in chemical exchange with respect to the NMR time scale, and, very probably, the major complex species has the C_3 -symmetry. In the case of the Zn^{2+} - H_3 notp^{tfe} system, a full complexation of the metal ion was expected on the basis of potentiometry. It was confirmed by the 19 F and 31 P NMR spectra (Fig. S23–S25†) where observation of only one symmetric signal points to the presence of only one enantiomeric pair of a single diastereoisomer (one combination of $\Delta/\Lambda + \delta/\lambda$

Dalton Transactions Paper

Table 5 Longitudinal relaxation times T_1 (ms) of the ¹⁹F NMR signals of H_3 notp^{tfe} and its complexes. Estimated deviation (based on repeated measurements) is generally < 5%

¹⁹ F Larmor frequency/MHz		565		376		282	
Temperature/°C	-						
Sample	δ(corr.)/ppm	$\frac{25}{T_1/\mathrm{ms}}$	37	25	37	25	37
H ₃ notp ^{tfe}	-57.12	0.66×10^{3}	0.85×10^{3}	0.97×10^{3}	1.2×10^{3}	1.1×10^{3}	1.5×10^{3}
[Mg ^{II} (notp ^{tfe})] ⁻ [Cr ^{III} (notp ^{tfe})]	-57.30	0.61×10^{3}	0.75×10^{3}	0.88×10^{3}	1.0×10^{3}	1.1×10^{3}	1.3×10^{3}
[Cr ^{III} (notp ^{tfe})]	-46.0	1.0^{a}	2.0^{a}	b	<u></u> b	<u></u> b	b
[Mn ^{II} (notp ^{tfé})]	-41.8	0.5^{a}	0.8^{a}	<u></u> b	<u></u> b	b	b
[Fe ^{III} (notp ^{tfe})]	-31.0	0.4^a	0.5^{a}	0.5^{a}	0.5^{a}	0.5^{a}	0.8^{a}
[Co ^{II} (notp ^{tfe})] ⁻	-50.5	44	57	64	71	78	83
[Ni ^{II} (notp ^{tfe})]	-48.5	3.2	3.9	3.3	4.0	3.6	4.8
[Cu ^{II} (notp ^{tfé})]	-54.2	4.7	5.7	4.1	5.0	3.8	5.2
[Zn ^{II} (notp ^{tfe})] ⁻	-57.21	0.61×10^3	0.76×10^{3}	0.95×10^3	1.1×10^3	1.0×10^3	1.4×10^3

^a Only a rough estimate due to the very fast relaxation. ^b Spectra cannot be successfully phased.

Table 6 Transversal relaxation times T2 (ms, diamagnetic compounds, measured with the CPMG sequence) and effective transversal relaxation times T_2^* (ms, paramagnetic compounds, calculated from the half-widths) of the ¹⁹F NMR signals of H_3 notp^{tfe} and its studied complexes. Estimated deviation (based on repeated measurements) is generally < 5%

¹⁹ F Larmor frequency/MHz		565	565		376		282	
Temperature/°C								
Sample	δ(corr.)/ppm	$\frac{25}{T_2/\mathrm{ms}}$	37	25	37	25	37	
H ₃ notp ^{tfe} [Mg ^{II} (notp ^{tfe})] ⁻ [Zn ^{II} (notp ^{tfe})] ⁻	-57.12 -57.30 -57.21	0.52×10^{3} 0.22×10^{3} 0.23×10^{3}	0.69×10^{3} 0.17×10^{3} 0.35×10^{3}	0.80×10^{3} 0.48×10^{3} 0.41×10^{3}	0.95×10^{3} 0.44×10^{3} 0.52×10^{3}	a a a	a a a	
$ \begin{aligned} & \left[\text{Cr}^{\text{III}}(\text{notp}^{\text{tfe}}) \right] \\ & \left[\text{Mn}^{\text{II}}(\text{notp}^{\text{tfe}}) \right]^{-} \\ & \left[\text{Fe}^{\text{III}}(\text{notp}^{\text{tfe}}) \right] \\ & \left[\text{Co}^{\text{II}}(\text{notp}^{\text{tfe}}) \right]^{-} \\ & \left[\text{Ni}^{\text{II}}(\text{notp}^{\text{tfe}}) \right]^{-} \\ & \left[\text{Cu}^{\text{II}}(\text{notp}^{\text{tfe}}) \right]^{-} \end{aligned} $	-46.0 -41.8 -31.0 -50.5 -48.5 -54.2	T_2^*/ms 0.22^b 0.08^b 0.21^b 1.8 1.9	0.17^{b} 0.07^{b} 0.26^{b} 1.0 $2.$ 2.6	$-c$ $-c$ 0.22^{b} 2.1 1.8	$ \begin{array}{c} -c \\ -c \\ 0.27^{b} \\ 1.3 \\ 2.6 \\ 2.2 \end{array} $	$-c \\ -c \\ 0.24^{b} \\ 2.3 \\ 2.3 \\ 1.7$	$-\frac{c}{c}$ 0.30 ^b 1.6 2.7 2.2	

^a Measurement of T_2 was not possible on a Varian VNMRS300 with an accessible probe. ^b Only a rough estimate due to the extreme broadness of the signal. ^c Spectra cannot be successfully phased.

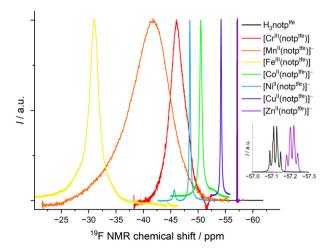


Fig. 11 Visual comparison of ¹⁹F NMR spectra of the studied transition metal complexes of H₃notp^{tfe} and free ligand. The inset shows spectra of diamagnetic H₃notp^{tfe} and [Zn(notp^{tfe})]⁻.

+ R/S) in the solution; it is very probably the octahedral $\Delta\lambda$ -SSS/ $\Lambda\delta$ -RRR species found in the solid state (see above). However, the ¹H and ¹³C{¹H} NMR spectra showed a fast fluxionality of the complex species: only two unresolved broad signals were found in the 1H NMR spectrum at 25 °C (Fig. S21†). The signals remained broad even at 5 °C which precludes further study. In the ¹³C{¹H} NMR spectrum, two very broad signals of the macrocyclic carbon atoms were found but other carbon atoms showed well-resolved sharp signals (Fig. S22†).

For the Cr3+, Mn2+ and Fe3+ complexes, only 19F NMR signals can be detected; signals of the other nuclei fully vanished, probably due to the relatively slow electronic relaxation caused by the symmetric electronic state of the metal ions $(t_{2g})^3$ for Cr3+, HS-d5 for the others). The 19F NMR signals of $[Cr^{III}(notp^{tfe})]$ and $[Mn^{II}(notp^{tfe})]^{-}$ are extremely broad $(\nu_{\frac{1}{3}} \approx 2.0)$ and ≈4.6 kHz at 565 MHz, respectively) whereas the ¹⁹F NMR signal of the [Fe^{III}(notp^{tfe})] complex is more narrow ($\nu_{\frac{1}{2}} \approx 1.1$

kHz at 565 MHz) and can be easily measured. However, the longitudinal relaxation of the 19 F NMR signals of all these complexes is extremely fast ($T_1 < 1$ ms, Tables 5 and 6).

In contrast to the paramagnetic complexes discussed above, all ¹H, ¹³C, ¹⁹F and ³¹P NMR spectra could be observed for the [Co^{II}(notp^{tfe})]⁻ complex. As several geometries of the complex were found in the solid state (see above), some dynamic equilibrium between the arrangements can be expected in the solution. However, there is only one symmetric (although broad) peak in the ¹⁹F/³¹P NMR spectra and only one set of a ¹H/¹³C {1H} signal. Thus, fluxionality of the [Co^{II}(notp^{tfe})] complex species is probably very fast. In the ¹H NMR spectra spreading over 200 ppm, 7 of the expected 8 signals were found (Fig. 12 and S13†) as the last signal was very close to the signal of the solvent (water/HDO) and overlapped; it was found by measuring the temperature dependence of the spectra (Fig. S13†). In the ¹³C{¹H} NMR spectra, all of the expected 5 signals were observed in the range from -500 to +200 ppm (Fig. 12 and S14†). A similar very large spectral ¹³C NMR range was observed previously for the Co^{2+} complex of the N-(2,2,2-trifluoroethyl) cyclam derivative.45

In the ¹⁹F NMR spectrum of [Co^{II}(notp^{tfe})]⁻, one broad signal centred at -50.5 ppm is present (Fig. S15†) which shows optimally fast longitudinal relaxation in order tens of milliseconds (T_1 40-80 ms, dependent on the external magnetic field strength and temperature, Table 5), although its transversal relaxation is very fast (Table 6). This compound also shows a broad $^{31}P\{^{1}H\}$ NMR signal at ca. 200 ppm (Fig. S16†). When the [Co^{II}(notp^{tfe})]⁻ complex is oxidized with K₂S₂O₈ or H₂O₂, a set of multiplets (pseudo-quartets due to similar values of J_{FH} and J_{FP}) of $[\text{Co}^{\text{III}}(\text{notp}^{\text{tfe}})]$ gradually appears in the 19F NMR spectrum (Fig. 13). It is consistent with the data from spectro-electrochemical experiments (Fig. S33†). The signals are narrow and their relaxation is slow. Thus, the formed [Co^{III}(notp^{tfe})] complex is obviously diamagnetic with a low-spin d^6 arrangement. Several close and hardly separable ¹⁹F NMR signals of [Co^{III}(notp^{tfe})] appear which can be explained by the presence of isomers with different R/S configurations on the phosphinate groups. These isomers are not exchanged due to kinetic inertness and non-fluxionality of the Co³⁺ complexes, and each affords individual signal(s).

The ¹⁹F NMR spectrum of the [Ni^{II}(notp^{tfe})]⁻ complex contains two broad ¹⁹F NMR signals separated by *ca.* 3 ppm, with

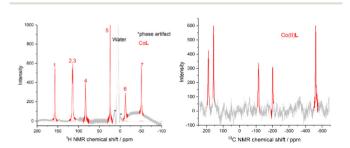


Fig. 12 1 H (left) and 13 C(1 H) (right) NMR spectra of the [Co II (notp tfe)]⁻ complex (D₂O, pD 7.4, 37 $^{\circ}$ C).

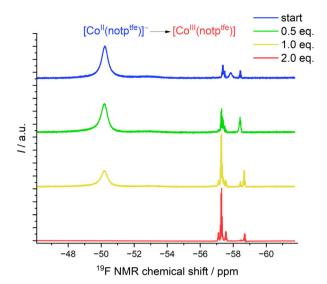


Fig. 13 19 F NMR spectra of the reaction mixtures showing gradual oxidation of $[Co^{II}(notp^{tfe})]^-$ to $[Co^{III}(notp^{tfe})]$ with $K_2S_2O_8$.

a relative intensity ca. 10:90% (Fig. S18†). In the case of $[Cu^{II}(notp^{tfe})]^-$, only one broad signal was observed (Fig. S20†). The signals of both complexes relax very fast with T_1 in the range of ca. 3–5 ms and T_2 ca. 2 ms (Tables 5 and 6).

Overall, the observed relaxation times are consistent with those found previously for the complexes of the related cyclam-based ligand 1,8-H₂te2p^{tfe} (Fig. 1) with the same pendant arm.⁴⁷ Fast movement of the 2,2,2-trifluoroethyl group effectively averages the distances; however, the mean value cannot be reliably calculated as it strongly correlates with electronic relaxation times whose exact values are not known and, for individual metal ions, can cover a wide range.47 However, the [Cr^{III}(notp^{tfe})], [Mn^{II}(notp^{tfe})] and [Fe^{III}(notp^{tfe})] complexes cannot be utilised in ¹⁹F MRI as they show very broad signals which relax too fast and, therefore, are not reliably detectable. The relaxation characteristics of the [Ni^{II}(notp^{tfe})] and [Cu^{II}(notp^{tfe})] complexes are more suitable but their potential use in the imaging experiments would need special ultrafast measurement techniques 90,91 due to too short relaxation times (few milliseconds). In this respect, the most promising is the [Co^{II}(notp^{tfe})] complex showing optimally fast longitudinal relaxation (T_1 in order of tens of milliseconds) which is suitable for standard MRI hardware. Furthermore, the redox potential of the [Co^{III}(notp^{tfe})]/ [Co^{II}(notp^{tfe})] pair has physiologically relevant value making the compound potentially employable as a redox probe.

Conclusions

A new 1,4,7-triazacyclononane-based ligand substituted with three (2,2,2-trifluoroethyl)phosphinate pendant arms was studied with respect to its potential use in contrast agents for ¹⁹F Magnetic Resonance Imaging (¹⁹F MRI). With a large metal ion Mn²⁺, the ligand forms complexes with twisted-trigonal-

antiprismatic geometry, whereas with the smaller ions Ni²⁺, Cu²⁺ and Zn²⁺, complex species with an octahedral geometry are formed. With the Co2+ ion, both environments were observed. Complexes of Co2+, Ni2+, Cu2+ and Zn2+ are very stable and are fully formed at pH > ~3. Paramagnetic metal ion complexes show very short relaxation times of the ¹⁹F NMR signal due to a short distance between the paramagnetic metal ion and the fluorine atoms (values found in the crystal structures are $\sim 5.5-6.5$ Å). Most of the studied complexes show too fast longitudinal relaxation for the application $(T_1 \text{ of the } Cr^{3+},$ Mn²⁺ and Fe³⁺ complexes in the sub-millisecond range; the Ni^{2+} and Cu^{2+} complexes ~4 ms), but T_1 of $[Co^{II}(notp^{tfe})]^{-}$ (40-80 ms) falls in a range suitable for easy exploitation. Electrochemical studies revealed the formation of a stable diamagnetic Co³⁺ complex ($E_{\frac{1}{2}}$ 0.95 V vs. SCE). Together with a suitable relaxation time, it makes the [Co^{III}(notp^{tfe})]/ [Co^{II}(notp^{tfe})] pair potentially useful as a smart redox-responsive contrast agent in 19F MRI.

Author contributions

Dalton Transactions

FK – investigation, methodology, validation, visualization, and writing – original draft; TD – investigation and writing – original draft, JK – conceptualization, investigation, methodology, validation, visualization, and writing – original draft, review and editing, IC – investigation, JH – investigation, AL – investigation, VK – investigation, PH – conceptualization, funding acquisition, and writing – review and editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The financial support of the Czech Science Foundation (GAČR) 22-34083S is acknowledged.

References

- 1 The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging, ed. A. E. Merbach, L. Helm and É. Tóth, John Wiley & Sons, 2nd edn, 2013.
- 2 C. S. Bonnet and É. Tóth, *Chimia*, 2016, 70, 102–108.
- 3 Y. A. Pirogov, Phys. Procedia, 2016, 82, 3-7.
- 4 I. Tirotta, V. Dichiarante, C. Pigliacelli, G. Cavallo, G. Terraneo, F. B. Bombelli, P. Metrangolo and G. Resnati, *Chem. Rev.*, 2015, 115, 1106–1129.
- 5 J. Ruiz-Cabello, B. P. Barnetta, P. A. Bottomley and J. W. M. Bulte, *NMR Biomed.*, 2011, 24, 114–129.
- 6 E. T. Ahrens, R. Flores, H. Xu and P. A. Morel, *Nat. Biotechnol.*, 2005, 23, 983–987.

- 7 T. Zhang, Q. Zhang, J.-H. Tian, J.-F. Xing, W. Guo and X.-J. Liang, MRS Commun., 2018, 8, 303–313.
- 8 K. Waxman, Ann. Emerg. Med., 1986, 15, 1423-1424.
- 9 A. R. Burgan, W. C. Herrick, D. M. Long and D. C. Long, *Biomater.*, *Artif. Cells*, *Artif. Organs*, 1988, 16, 681–682.
- 10 K. C. Lowe, Blood Rev., 1999, 13, 171-184.
- 11 B. Remy, G. Deby-Dupont and M. Lamy, *Br. Med. Bull.*, 1999, 55, 277–298.
- 12 G. A. Peyman, J. A. Schulman and B. Sullivan, Surv. Ophthalmol., 1995, 39, 375–395.
- 13 R. D. Bourke, R. N. Simpson, R. J. Cooling and J. R. Sparrow, *Arch. Ophthalmol.*, 1996, 114, 537–544.
- 14 J. Pastor, R. M. Coco, I. Fernandez-Bueno, M. L. Alonso-Alonso, J. Medina, A. Sanz-Arranz, F. Rull, M. J. Gayoso, A. Dueñas, M. T. Garcia-Gutierrez, L. Gonzalez-Buendia, S. Delgado-Tirado, E. Abecia, M. Ruiz-Miguel, M. A. Serrano, J. M. Ruiz-Moreno and G. K. Srivastava, *Retina*, 2017, 37, 1140–1151.
- I. Georgalas, I. Ladas, I. Tservakis, S. Taliantzis,
 E. Gotzaridis, D. Papaconstantinou and C. Koutsandrea,
 Cutaneous Ocul. Toxicol., 2011, 30, 251–262.
- 16 C. Gatto, P. Ruzza, L. Giurgola, C. Honisch, O. Rossi, M. R. Romano, E. Ragazzi and J. D'Amato Tóthová, ACS Omega, 2023, 8, 365–372.
- 17 T. Stahl, D. Mattern and H. Brunn, *Environ. Sci. Eur.*, 2011, 23, 38.
- 18 P. Hermann, J. Blahut, J. Kotek and V. Herynek, *Met. Ions Life Sci.*, 2021, **22**, 239–270.
- 19 P. K. Senanayake, A. M. Kenwright, D. Parker and S. K. van der Hoorn, *Chem. Commun.*, 2007, 2923–2925.
- 20 A. M. Kenwright, I. Kuprov, E. De Luca, D. Parker, S. U. Pandya, P. K. Senanayake and D. G. Smith, *Chem. Commun.*, 2008, 2514–2516.
- 21 K. H. Chalmers, E. De Luca, N. H. M. Hogg, A. M. Kenwright, I. Kuprov, D. Parker, M. Botta, J. I. Wilson and A. M. Blamire, *Chem. Eur. J.*, 2010, **16**, 134–148.
- 22 Z.-X. Jiang, Y. Feng and Y. B. Yu, Chem. Commun., 2011, 47, 7233–7235.
- 23 K. H. Chalmers, M. Botta and D. Parker, *Dalton Trans.*, 2011, 904–913.
- 24 P. Harvey, K. H. Chalmers, E. De Luca, A. Mishra and D. Parker, *Chem. Eur. J.*, 2012, **18**, 8748–8757.
- 25 P. Harvey, I. Kuprov and D. Parker, Eur. J. Inorg. Chem., 2012, 2015–2022.
- 26 M. P. Placidi, M. Botta, F. K. Kálmán, G. E. Hagberg, Z. Baranyai, A. Krenzer, A. K. Rogerson, I. Tóth, N. K. Logothetis and G. Angelowski, *Chem. - Eur. J.*, 2013, 19, 11644–11660.
- 27 N. Cakić, T. Savić, J. Stricker-Shaver, V. Truffault, C. Platas-Iglesias, C. Mirkes, R. Pohmann, K. Scheffler and G. Angelovski, *Chem. Commun.*, 2016, 52, 9224–9227.
- 28 K. Srivastava, E. A. Weitz, K. L. Peterson, M. Marjańska and V. C. Pierre, *Inorg. Chem.*, 2017, **56**, 1546–1557.
- 29 R. Pujales-Paradela, T. Savić, D. Esteban-Gómez, G. Angelovski, F. Carniato, M. Botta and C. Platas-Iglesias, Chem. – Eur. J., 2019, 25, 4782–4792.

30 R. Pujales-Paradela, T. Savić, P. Pérez-Lourido, D. Esteban-Gómez, G. Angelovski, M. Botta and C. Platas-Iglesias, Inorg. Chem., 2019, 58, 7571–7583.

- 31 V. Herynek, M. Martinisková, Y. Bobrova, A. Gálisová, J. Kotek, P. Hermann, F. Koucký, D. Jirák and M. Hájek, *Magn. Reson. Mater. Phys., Biol. Med.*, 2019, **32**, 115–122.
- 32 R. B. Lauffer, Chem. Rev., 1987, 87, 901-927.
- 33 D. Xie, M. Yu, R. T. Kadakia and E. L. Que, *Acc. Chem. Res.*, 2020, 53, 2–10.
- 34 K. Srivastava, G. Ferrauto, V. G. Young Jr., S. Aime and V. C. Pierre, *Inorg. Chem.*, 2017, **56**, 12206–12213.
- 35 M. Yu, B. S. Bouley, D. Xie and E. L. Que, *Dalton Trans.*, 2019, 48, 9337–9341.
- 36 NIST Standard Reference Database 46 (Critically Selected Stability Constants of Metal Complexes), Version 7.0, 2003, distributed by NIST standard Reference Data, Gaithersburg, MD 20899, USA.
- 37 M. Yu, D. Xie, K. P. Phan, J. S. Enriquez, J. J. Luci and E. L. Que, *Chem. Commun.*, 2016, 52, 13885–13888.
- 38 A. E. Thorarinsdottir, A. I. Gaudette and T. D. Harris, *Chem. Sci.*, 2017, **8**, 2448–2456.
- 39 A. I. Gaudette, A. E. Thorarinsdottir and T. D. Harris, *Chem. Commun.*, 2017, 53, 12962–12965.
- 40 M. Yu, B. S. Bouley, D. Xie, J. S. Enriquez and E. L. Que, J. Am. Chem. Soc., 2018, 140, 10546–10552.
- 41 J. Blahut, P. Hermann, A. Gálisová, V. Herynek, I. Císařová, Z. Tošner and J. Kotek, *Dalton Trans.*, 2016, 45, 474–478.
- 42 J. Blahut, K. Bernášek, A. Gálisová, V. Herynek, I. Císařová, J. Kotek, J. Lang, S. Matějková and P. Hermann, *Inorg. Chem.*, 2017, 56, 13337–13348.
- 43 R. Pujales-Paradela, T. Savić, I. Brandariz, P. Pérez-Lourido, G. Angelovski, D. Esteban-Gómez and C. Platas-Iglesias, Chem. Commun., 2019, 55, 4115–4118.
- 44 D. Xie, L. E. Ohman and E. L. Que, *Magn. Reson. Mater. Phys., Biol. Med.*, 2019, 32, 89-96.
- 45 J. Blahut, L. Benda, J. Kotek, G. Pintacuda and P. Hermann, *Inorg. Chem.*, 2020, **59**, 10071–10082.
- 46 Z. Kotková, F. Koucký, J. Kotek, I. Císařová, D. Parker and P. Hermann, *Dalton Trans.*, 2023, **52**, 1861–1875.
- 47 F. Koucký, J. Kotek, I. Císařová, J. Havlíčková, V. Kubíček and P. Hermann, *Dalton Trans.*, 2023, **52**, 12208–12223.
- 48 W. L. F. Armarego and C. L. L. Chai, *Purification of Laboratory Chemicals*, *Butterworth Heinemann*, An imprint of Elsevier Science, 5th edn, 2003.
- 49 T. M. Gøgsig, L. S. Søbjerg, A. T. Lindhardt, K. L. Jensen and T. Skrydstrup, *J. Org. Chem.*, 2008, 73, 3404–3410.
- 50 E. T. McBee, D. H. Campbell and C. W. Roberts, *J. Am. Chem. Soc.*, 1955, 77, 3149–3151.
- 51 S. Xia, Y.-B. Shi, F.-F. He and H.-B. Wang, Acta Crystallogr., Sect. E: Struct. Rep. Online, 2010, 66, 02779.
- 52 K. Wieghardt, U. Bossek, P. Chaudhuri, W. Herrmann, B. C. Menke and J. Weiss, *Inorg. Chem.*, 1982, **21**, 4308–4314.
- 53 M. Krejčik, M. Daněk and J. Hartl, *J. Electroanal. Chem.*, 1991, 317, 179–187.
- 54 M. Försterová, I. Svobodová, P. Lubal, P. Táborský, J. Kotek, P. Hermann and I. Lukeš, *Dalton Trans.*, 2007, 535–549.

- 55 C. F. Baes Jr. and R. E. Mesmer, The Hydrolysis of Cations, Wiley, New York, 1976.
- 56 M. Kývala and I. Lukeš, International conference, chemometrics × 95, abstract book p. 63, Pardubice (Czech Republic), 1995; full version of TMOPIUM..
- 57 L. Krause, R. Herbst-Irmer, G. M. Sheldrick and D. Stalke, J. Appl. Crystallogr., 2015, 48, 3–10.
- 58 (a) G. M. Sheldrick, SHELXT2014/5. Program for Crystal Structure Solution from Diffraction Data, University of Göttingen, Göttingen, 2014; (b) G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 2008, 64, 112–122.
- 59 (a) C. B. Hübschle, G. M. Sheldrick and B. Dittrich, ShelXle:
 a Qt graphical user interface for SHELXL, University of Göttingen, Göttingen, 2014; (b) C. B. Hübschle, G. M. Sheldrick and B. Dittrich, J. Appl. Crystallogr., 2011, 44, 1281–1284; (c) G. M. Sheldrick, Acta Crystallogr., Sect. C: Struct. Chem., 2015, 71, 3–8; (d) G. M. Sheldrick, SHELXL-2017/1. Program for Crystal Structure Refinement from Diffraction Data, University of Göttingen, Göttingen, 2017.
- 60 K. Bazakas and I. Lukeš, Dalton Trans., 1995, 1133-1137.
- 61 J. Šimeček, M. Schulz, J. Notni, J. Plutnar, V. Kubíček, J. Havlíčková and P. Hermann, *Inorg. Chem.*, 2012, **51**, 577–590.
- 62 J. Huskens and A. D. Sherry, J. Am. Chem. Soc., 1996, 118, 4396-4404.
- 63 I. Lukeš, J. Kotek, P. Vojtíšek and P. Hermann, Coord. Chem. Rev., 2001, 216–217, 287–312.
- 64 B. Drahoš, V. Kubíček, C. S. Bonnet, P. Hermann, I. Lukeš and E. Tóth, *Dalton Trans.*, 2011, 40, 1945–1951.
- 65 V. Kubíček, Z. Böhmová, R. Ševčíková, J. Vaněk, P. Lubal, Z. Poláková, R. Michalicová, J. Kotek and P. Hermann, *Inorg. Chem.*, 2018, 57, 3061–3072.
- 66 C. F. G. C. Geraldes, A. D. Sherry and W. P. Cacheris, *Inorg. Chem.*, 1989, 28, 3336–3341.
- 67 J. Gao, H. He, W. Yang, J.-G. Hou and J. Kang, *Lab. Rob. Autom.*, 1998, **10**, 229–233.
- 68 M. I. Kabachnik, T. Ya. Medved, Yu. M. Polikarpov, B. K. Tscherbakov, F. I. Belskij, E. I. Mamrosov and M. P. Pasechnik, *Izv. Akad. Nauk SSSR*, *Ser. Khim.*, 1984, 835–843.
- 69 H. F. Bauer and W. C. Drinkard, J. Am. Chem. Soc., 1960, 82, 5031-5032.
- 70 D. F. Evans, J. Chem. Soc., 1959, 2003-2005.
- 71 D. M. Corsi, C. Platas-Iglesias, H. van Bekkum and J. A. Peters, *Magn. Reson. Chem.*, 2001, **39**, 723–726.
- 72 T. L. Hatfield, R. J. Staples and D. T. Pierce, *Inorg. Chem.*, 2010, 49, 9312–9320.
- 73 S. Aime, A. S. Batsanov, M. Botta, J. A. K. Howard, D. Parker, K. Senanayake and G. Williams, *Inorg. Chem.*, 1994, 33, 4696–4706.
- 74 S. Aime, M. Botta, M. Fasano, M. P. M. Marques, C. F. G. C. Geraldes, D. Pubanz and A. E. Merbach, *Inorg. Chem.*, 1997, 36, 2059–2068.
- 75 S. Aime, A. Barge, F. Benetollo, G. Bombieri, M. Botta and F. Uggeri, *Inorg. Chem.*, 1997, 36, 4287–4289.

76 F. Benetollo, G. Bombieri, L. Calabi, S. Aime and M. Botta, *Inorg. Chem.*, 2003, 42, 148–157.

Dalton Transactions

- 77 M. Woods, M. Botta, S. Avedano, J. Wang and A. D. Sherry, Dalton Trans., 2005, 3829–3837.
- 78 P. Vojtíšek, P. Cígler, J. Kotek, J. Rudovský, P. Hermann and I. Lukeš, *Inorg. Chem.*, 2005, 44, 5591–5599.
- 79 J. Kotek, J. Rudovský, P. Hermann and I. Lukeš, *Inorg. Chem.*, 2006, 45, 3097–3102.
- 80 P. Urbanovský, J. Kotek, I. Císařová and P. Hermann, *Dalton Trans.*, 2020, **49**, 1555–1569.
- 81 W. D. Kim, G. E. Kiefer, J. Huskens and A. D. Sherry, *Inorg. Chem.*, 1997, **36**, 4128–4134.
- 82 J. Rohovec, I. Lukeš and P. Hermann, *New J. Chem.*, 1999, 23, 1129–1132.
- 83 Z. Kotková, G. A. Pereira, K. Djanashvili, J. Kotek, J. Rudovský, P. Hermann, L. Vander Elst, R. N. Muller, C. F. G. C. Geraldes, I. Lukeš and J. A. Peters, *Eur. J. Inorg. Chem.*, 2009, 119–136.

- 84 Y.-H. Su, S.-S. Bao and L.-M. Zheng, *Inorg. Chem.*, 2014, 53, 6042–6047.
- 85 M. I. Kabachnik, M. Y. Antipin, B. K. Tscherbakov, A. P. Baranov, Yu. T. Struchkov, T. Y. Medved and Yu. M. Polikarpov, *Koord. Khim.*, 1988, 14, 536–542.
- 86 E. Cole, D. Parker, G. Ferguson, J. F. Gallagher and B. Kaitner, *J. Chem. Soc., Chem. Commun.*, 1991, 1473–1475.
- 87 E. Cole, R. C. B. Copley, J. A. K. Howard, D. Parker, G. Ferguson, J. F. Gallagher, B. Kaitner, A. Harrison and L. Royle, J. Chem. Soc., Dalton Trans., 1994, 1619–1629.
- 88 S.-S. Bao, G.-S. Chen, Y. Wang, Y.-Z. Li, L.-M. Zheng and Q.-H. Luo, *Inorg. Chem.*, 2006, 45, 1124–1129.
- 89 M. Zalewski, D. Janasik, A. Wierzbicka and T. Krawczyk, *Inorg. Chem.*, 2022, 61, 19524–19542.
- K. H. Chalmers, A. M. Kenwright, D. Parker and A. M. Blamire, *Magn. Reson. Med.*, 2011, 66, 931–936.
- 91 F. Schmid, C. Höltke, D. Parker and C. Faber, *Magn. Reson. Med.*, 2013, **69**, 1056–1062.